Magnitude of Drug-Resistant Enterococcus Species from intestinal Tracts of Hospitalized Pediatric Patients in Debreberhan Referral Hospital, Debreberhan, Ethiopia

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Abstract

Background: Multi-drug-resistant enterococci are the major source of infection as well as nosocomial spread. There is scarcity of data on drug-resistant enterococci in developing country including Ethiopia. Therefore, this study aimed to determine the magnitude of drug resistant enterococcus species from intestinal tracts of hospitalized pediatric patients.

Method: The study was conducted among hospitalized pediatric patients at DebreBerhan referral hospital, from February 15 to March, 25 2016. Rectal swabs was collected and processed for bacterial isolation and susceptibility testing. The isolates was identified to species level by cultural characteristics, Gram’s stain, catalase test and other biochemical tests. Susceptibility testing to antimicrobial agents was done using Kirby–Bauer disk diffusion method.

Result: Enterococci were isolated from 12 (23%) of the study participants. The isolates were Enterococcus faecium(50%), Enterococcus faecalis(33.3%) and Enterococcus gallinarum(16.7%). Among 12 tested Enterococci isolates, 5 (41.7%) were resistant to ampicillin, 7(58.3%) to streptomycin, 6 (50%) to gentamycin, 7(58.3%) to ciprofloxacin, 5(41.7%) to norfloxacin and 8(66.7%) to erythromycin. Multiple drugresistance was observed among 75% of E. faecium and E. faecalis. Vancomycin resistant Enterococci were observed in 16.7% of E. faeicium isolates.

Conclusion: This study reveals high rate of fecal colonization by multidrug-resistant enterococci and prevalence of vancomycin resistance strains. Thus periodic surveillance of antibiotic susceptibilities is recommended to detect emerging resistance and to prevent its spread.

Keywords: Magnitude, Drug-resistance, Enterococcus species, Pediatric Patients

1. INTRODUCTIONS

Enterococci are normal inhabitant of the gastrointestinal tract. However, they can also be significant pathogens causing several infections. The most common nosocomial infections caused by these organism are urinary tract infections (associated with instrumentations and antimicrobials administration) (1). The emergence of vancomycin resistance enterococci (VRE) is a global issue due to few option left for disease management. Besides drug resistant Enterococci can colonize the intestinal tract of hospitalized patients and become major source of infection as well as nosocomial spread (2, 3).

In humans, enterococcal infections may be caused by at least 12 species but most clinical infections are due to either Enterococcus faecalis or E. faecium. E. faecalis is the most common cause (80–90%) followed by E. faecium (10–15%). Occasional infections are due to Enterococcus gallinarum, Enterococcus raffinosus, Enterococcus casseliflavus, Enterococcus avium, Enterococcus pseudaoavium, Enterococcus malodoratus, Enterococcus mundtii, Enterococcus durans, and Enterococcus hirae (4). The proportion of isolates of motile Enterococci (E. gallinarum, E. casseliflavus) is low. But they are intrinsically resistant to vancomycin and inappropriate treatment may contribute to morbidity and mortality (5).
Several studies have documented that enterococcal infections are most commonly caused by the patient’s own commensal flora. Colonization may occur long before or immediately before infection, but it plays a major role in the development of nosocomial infection (2). Despite the importance of these etiologic agents there is a dearth of information regarding antimicrobial resistance of Enterococcus species isolated from intestinal tract of hospitalized patients in Ethiopia. Thus, the present study will be conducted to determine antimicrobial resistance pattern of fecal enterococci isolates from hospitalized pediatric patients.

2. METHODS

2.1. Study Area and Periods
The study was conducted at Debre Berhan referral hospital, from February 15, 2016 to March 25, 2016. It is located 130 kms Northeast of Addis Ababa.

2.2. Study Design
Cross sectional study was conducted and the study participant was recruited conveniently.

2.3. Sampling Technique and Sample Size
52 patients, who had at least 2 days of hospital stay at pediatric ward of Debre Berhan referral hospital were enrolled by using convenience sampling technique.

2.4. Inclusion and Exclusion Criteria
Pediatric patients aged 0 to 15 years and having at least 48 hour hospital stay during the study period and fulfill the inclusion criteria was included while who cannot respond to the interview and for children without getting permission from guardian and consent was excluded.

2.5. Data Collection Techniques
Scio-demographic data was collected by pretested questionnaires after obtaining a written informed consent from study participants. Fecal samples were collected in sterile plastic stool containers. From critically ill patients rectal swabs were collected using sterile cotton swab moistened in sterile normal saline solution. Then, the swabs were immersed in well-labeled Cary-Blair semi-solid medium prepared in screw-capped tubes and transferred to the Debre Berhan university medical microbiology laboratory.

2.1.1. Culture and Identification
Stool specimens and rectal swabs were inoculated onto Bile Esculinazide agar plates with and without 6 µg/ml of vancomycin and incubated at 37°C for 24 h. colonies with colourless or grey and surrounded by a black halo (hydrolysis of esculin) were sub-cultured and identified as Enterococci by additional tests like gram stain, catalase test, 6.5% NaCl test, growth at 45°C and motility test as recommended by Facklam and Collins (6), Manero and Blanch (7). Identification of these isolates to species level was performed by API-20 Streptococcus system (bioMe´rieux).

2.1.2. Antibiotic Susceptibility Testing
Antimicrobial susceptibility studies were performed by disc diffusion (Kirby–Bauer) method according to Clinical Laboratory Standards Institute (CLSI) for widely used drugs in Ethiopia with their respective dose. Minimum inhibitory concentrations (MICs) for vancomycin were determined using E-test strips.

2.6. Data Processing, Analysis
The collected data was clearly summarized, filled and analysed by using SPSS version 21. Descriptive statistics was employed to examine the finding, and the result was presented by using tables, charts and graphs. P-value less than or equal to 0.05 was considered as statistically significant value.

2.7. Quality Assurance
The questionnaires was pretested a week before actual data collection. The quality of reagents and equipment was checked and used according to manufacturer directions. The data was collected by trained data collectors and the result was recorded carefully and correctly. Standard operating procedure was applied during specimen collection, culture, drug susceptibility test and biochemical test. E. faecalis ATCC 29212 was used as a quality control strain for performing antimicrobial tests.
2.8. Ethical Consideration
The study protocol was reviewed and approved by the ethical and review committee of Debre Berhan university. For all study participant the objective of the study was explained and written informed consent was obtained. Those positive for enterococcus was referred to their respective clinicians for further management.

3. RESULT

3.1. Demographic Characteristics
Among 52 participants, 27 (52%) were males and 25(48%) were females. The mean age of the patient’s was 3 years. 78.8% study participant had a history of exposure to one or more antimicrobial agent in the last 2 weeks and 21.2% were without exposure and the average hospital stay was 17.5 days with a range of 2-45 days (Table 1).

3.2. Enterococci Isolates
Among all participants, 12 (23%) of the study participants were positive for at least one Enterococcus spp. There was no statistically significant association between isolation of Enterococci with age, sex, hospital duration and antibiotic history (Table 1).

Table 1. Socio demographic characteristics and Enterococcus culture positivity at DebreBerhan Referral Hospital, DebreBerhan, Ethiopia

<table>
<thead>
<tr>
<th>Variable</th>
<th>CulturePositive n (%)</th>
<th>CultureNegative n (%)</th>
<th>Total (%)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age category in year</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0-7 year</td>
<td>8(25.8%)</td>
<td>23(74.2%)</td>
<td>31(59.6%)</td>
<td>0.432</td>
</tr>
<tr>
<td>8-15 year</td>
<td>4(19%)</td>
<td>17(81%)</td>
<td>21(40.4%)</td>
<td></td>
</tr>
<tr>
<td>Sex</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>7(26%)</td>
<td>20(74%)</td>
<td>27(52%)</td>
<td>0.546</td>
</tr>
<tr>
<td>Female</td>
<td>5(20%)</td>
<td>20(80%)</td>
<td>25(48%)</td>
<td></td>
</tr>
<tr>
<td>Hospital duration</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2–15 day</td>
<td>3(10.7%)</td>
<td>25(89.3%)</td>
<td>28(53.8%)</td>
<td>0.135</td>
</tr>
<tr>
<td>&gt;15 day</td>
<td>9(37.5%)</td>
<td>15(62.5%)</td>
<td>24(46.2%)</td>
<td></td>
</tr>
<tr>
<td>Previous antibiotic treatment</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Yes</td>
<td>7(17.1%)</td>
<td>34(82.9%)</td>
<td>41(78.8%)</td>
<td>0.313</td>
</tr>
<tr>
<td>No</td>
<td>5(45.5%)</td>
<td>6(54.5%)</td>
<td>11(21.2%)</td>
<td></td>
</tr>
</tbody>
</table>

3.3. Species Distribution
The distribution of species is that a total of 12 enterococcal isolates were obtained from 52 patients. The commonly enterococcal isolates were E. faecium(50%) followed by E. faecalis(33.3%) and E. gallinarum(16.7%) (Table 2).

Table 2. Distribution of Enterococcus species isolated from intestinal tract of hospitalized patients in Debre Berhan Referral Hospital, DebreBerhan, Ethiopia.

<table>
<thead>
<tr>
<th>Enterococcus species</th>
<th>N (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>E. faecium</td>
<td>6 (50%)</td>
</tr>
<tr>
<td>E. faecalis</td>
<td>4 (33.3%)</td>
</tr>
<tr>
<td>E. gallinarum</td>
<td>2 (16.7%)</td>
</tr>
<tr>
<td>Total</td>
<td>12</td>
</tr>
</tbody>
</table>

3.4. Antimicrobial Resistance of Enterococcal Isolates

3.4.1. B-Lactam Resistance
1/4 (25%) E. faecalisand 4/6 (66.7%) E. faecium were resistant to ampicillin. AllE. fecalisand5/6 (83.3) E. faeciumisolates were resistant to penicillin. 1/2 (50%) occasional Enterococcus species (E. gallinarum) were resistant to penicillin.

3.4.2. Aminoglycoside Resistance
High-level resistance to gentamicin and streptomycin was detected by the high content disk. Gentamycin resistant were observed in 50 % of E. faecalisand 66.7 % of E. faecium. 2/4 (50%) E. faecalisand 66.7% E. faeciumwere resistant to streptomycin.
3.4.3. Vancomycin Resistant Enterococci

E. faecalis isolates was not resistance to vancomycin while 1 (16.7%) of E. faecium isolates were resistant to vancomycin (Table-3).

### Table 3. Antibiotic resistance profile of Enterococcus species at DebreBerhan Referral Hospital, DebreBerhan, Ethiopia

<table>
<thead>
<tr>
<th>Antibiotics</th>
<th>Resistant isolates (%)</th>
<th>E. faecalis(n,4)</th>
<th>E. faecium(n,6)</th>
<th>Other species(n,2)</th>
<th>Total(n,12)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ampicillin (10 μg)</td>
<td>1(25%)</td>
<td>4(66.7%)</td>
<td>0(0%)</td>
<td>5(41.7%)</td>
<td></td>
</tr>
<tr>
<td>Penicillin (10 μg)</td>
<td>4(100%)</td>
<td>5(83.3%)</td>
<td>1(50%)</td>
<td>10(83.3%)</td>
<td></td>
</tr>
<tr>
<td>Gentamicin (120 μg)</td>
<td>2(50%)</td>
<td>4(66.7%)</td>
<td>0(0%)</td>
<td>6(50%)</td>
<td></td>
</tr>
<tr>
<td>Ciprofloxacin (5 μg)</td>
<td>3(75%)</td>
<td>4(66.7%)</td>
<td>0(0%)</td>
<td>7(58.3%)</td>
<td></td>
</tr>
<tr>
<td>Streptomycin (300 μg)</td>
<td>2(50%)</td>
<td>4(66.7%)</td>
<td>1(50%)</td>
<td>7(58.3%)</td>
<td></td>
</tr>
<tr>
<td>Erythromycin (15 μg)</td>
<td>1(25%)</td>
<td>5(83.3%)</td>
<td>2(100%)</td>
<td>8(66.7%)</td>
<td></td>
</tr>
<tr>
<td>Norfloxacin (30 μg)</td>
<td>1(25%)</td>
<td>3(50%)</td>
<td>1(50%)</td>
<td>5(41.7%)</td>
<td></td>
</tr>
<tr>
<td>Vancomycin (6 μg/ml)</td>
<td>0(0%)</td>
<td>1 (16.7%)</td>
<td>0(0%)</td>
<td>1(8.3%)</td>
<td></td>
</tr>
</tbody>
</table>

### 4. DISCUSSIONS

This study investigated the prevalence and antibacterial resistance patterns of enterococci isolated from fecal samples of hospitalized pediatrics patients. In this study the distribution of enterococcus isolates were E. faecium (50%), E. faecalis (33.3%) and E. gallinarum (16.7%). This is comparable to the study done in Brazil (25). But disagreement with reports from United States (8). In this study, the predominant enterococcus isolates was E. faecium. Study in Singapore has also reported an increase in E. faecium from 78.9% to 91.8% over a period of 5 years from 2006 to 2010 from clinical cultures (9). Another study from India has also reported 66% E. faecium from blood sample (10). In this study the prevalence of E. gallinarum was 16.7% which is higher than study done in Ethiopia (1). This might be indicate that, even though enterococci including E. gallinarum are infrequently isolated from clinical specimens, they have been implicated in a wide variety of invasive infections in humans, especially immune compromised or chronically ill patients.

In this study, E. faecalis isolates showed 25% resistance rate to ampicillin. It is higher than the resistance rates reported in Kuwait, Hong Kong and Brazil, which is 0-8.3% (11, 12, 13), and lower than 60.7% reported from Gaza (14). Resistance rates to ampicillin was observed in 66.7% of E. faecium isolates which is comparable with study done in Gaza 66.7% (14). However, lower than study reported from Israel (15). All E. faecalis and 83.3% E. faecium isolates were resistant to penicillin which is similar to study done in India from clinical isolates (10). The reason for higher prevalence of β-lactam antibiotic resistance in this study might be due to chronic cases and wider usage of broad spectrum antibiotics relative to enterococcus isolates possess an intrinsically relative resistance to penicillin and ampicillin. Furthermore, E. faeciumis less susceptible to β-lactam agents than E. faecalis because their penicillin-binding proteins (PBPs) have lower affinities for these antibiotics and some strains have plasmid-encoded β-lactamase.

Aminoglycosides are frequently used in combination with cell wall active antibiotics for severe enterococcal infections. Since enterococcal resistance to gentamicin and streptomycin occurs by different mechanisms, it is important to test susceptibility to both agents. Enterococci with high level resistance to streptomycin are susceptible to gentamicin. And also gentamicin resistance is a good predictor of resistance to other aminoglycosides except streptomycin (16). In this study, E. faecalis and E. faecium showed resistance for many drugs. Concomitant resistance of high level aminoglycoside resistance (HLAR) strains to the β-lactam antibiotic (ampicillin) was quite higher (25% of E. faecalis and 66.7% of E. faecium strains). This finding is a cause of concern, because the synergistic activity of the combination of β-lactam antibiotics with HLAR in the treatment of enterococcal infections is totally abolished. In such instances, controlling the spread of these organisms have supreme importance.

In this study 58.3% of enterococci isolate were resistant to ciprofloxacin and 41.7% of the isolates were resistant to norfloxacin. Other alternative antibiotics to treat infection by enterococcus also showed high rates of resistance (erythromycin (66.7% of resistance) and (58.3%) of resistance to streptomycin). The high rates of resistance in present study might be due to excessive or inappropriate use of those antibiotics for empirical treatment of mixed nosocomial infections caused by enterococci.
The emergence of VRE is also due to the inappropriate use of cephalosporin as well as poor hospital infection control measures. This study showed 16.7% E. faecium resistant to vancomycin which is higher than study done in Egypt 4% (17), Iran 6.2% (18), South Africa 10.2% (19) and report from Korea 12% (20) and lower than report from Turkey 34.8% (21). The possible reason for the emergence of VRE in this study might be due to antibiotic selective pressure because the patients had long duration in hospital and high rate of antibiotics treatment.

The increase of invasive infections caused by multi resistant E. faecium, however, did not only increase the total burden of nosocomial enterococcal infections, but also resulted in a partial replacement of E. faecalis by E. faecium as a cause of hospital-associated infections. Several studies showed that an increased proportion of nosocomial enterococcal infections caused by E. faecium.

5. CONCLUSIONS AND RECOMMENDATIONS

23% of the hospitalized pediatrics patients carried enterococci in their gastrointestinal tracts and E. faecium was the predominant species. E. faecium showed highest resistance rate to vancomycin. Therefore, regular monitoring for the presence of VRE in both hospitals and the community, effective strategies for the prevention of antimicrobial resistance should be practiced. Unnecessary use of antibiotics and ignorance infection control measure should be stopped.

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AUTHORS’ CONTRIBUTIONS

SG-performed the laboratory activities. BF, SG and TA-analyzed the data. TA-wrote the manuscript. All authors read and approved the final manuscript.

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