Utilization of Seaweeds to Enhance Growth and Nutritive Status of Amaranthus Caudatus L

T.Kumareswari

Ph.D scholar, Dept. of Botany St. Mary's College (Autonomous) Tuticorin, Tamil Nadu, India *jkumareswari@yahoo.com*

S. Maria Victorial Rani

Head of the Dept. of Botany St.Mary's College(Autonomous) Tuticorin, Tamil Nadu, India. *smvr1960@yahoo.com.in*

Abstract: Modern agriculture is looking for new biotechnological advances that would allow a reduction in the use of chemical inputs without affecting crop yield or the farmer's income. Seaweeds, marine organic resource was formulated into organic aqueous extract and was applied to Amaranthus caudatus L. Seaweed liquid fertilizer (SLF) of Padina pavonia enhanced the shoot length, root length and leaf area by 83%, 131% and 79% respectively under the soil application in comparison with control. Application of SLF of Padina tetrastromatica also increased number of leaves (56.8%), above ground biomass (145%), flavonoid (91.8%), total carbohydrate (61.6%), free radical scavenging activity (67.8%) and superoxide radical scavenging activity (175%) when compared to control. Application of SLF of Stoechospermum marginatum enhanced the total cholorophyll, total phenolic content, vitamin C, carotenoid, total soluble protein and free amino acid in relation to control. The study revealed that SLF of Padina pavonia was found to be highly effective in enhancing only the growth and leaf area. Application SLF of Padina tetrastromatica also enhancing number of leaves and above ground biomass. Even though, SLF of Stoechospermum marginatum did not promote any of these growth parameters, but stimulated the production of photosynthetic pigments, phytochemical and biochemical constituents. The results of the present study showed that SLF of three seaweeds maintained well the plant growth to increase yield of consumable product.

Keywords: Seaweed liquid fertilizer (SLF), Stoechospermum marginatum (Ag) Kulz, Padina pavonia (L.) J.V. Lamouroux, Padina tetrastromatica Hanck and Amaranthus caudatus L.

1. INTRODUCTION

Application of synthetic growth promoters in agriculture farming has been in practice for many decades. However, continuous use of inorganic fertilizers in traditional farming has been losing its popularity because of its adverse effect in altering the chemistry of the soil, making it less congenial for plant growth. Moreover, the harmful effects of inorganic fertilizers have serious impact on human health [1]. Seaweeds constitute one of the important biotic components of the ocean and might serve as an alternative to inorganic fertilizers [2]. The use of seaweed manure in conjunction with the inorganic fertilizers has been found to be better than the other organic input for the growth and development of plant [3]. Seaweed liquid fertilizers will be useful for achieving higher agricultural production, because the extract contains growth promoting hormones (IAA and IBA), cytokinins, gibbrellins, trace elements, vitamins, amino acids, antibiotics and micronutrients [4]. The higher amount of water soluble potash, other minerals and trace elements present in seaweeds are readily absorbed by plants and they control deficiency diseases [5]. The carbohydrates and other organic matter present in seaweeds alter the nature of soil and improve its moisture retaining capacity [5]. Liquid seaweed extract when applied to seed, soil or sprayed on crops increased seed germination percentage, nutrient uptake, growth [6] and yield of crops [7]. The diluted liquid seaweed extract also enhanced the plant's defence against diseases and increases salt [8] and biotic and abiotic stress [9, 10 and 11]. Considering the importance of seaweed extract for agricultural applications, an attempt has been to evaluate the growth enhancing properties of extracts prepared from three species of brown seaweeds namely: Padina pavonia, Padina tetrastromatica Hanck and Stoechospermum marginatum (Ag) Kulz on a green leafy vegetable plant (Amaranthus caudatus L.)

2. MATERIALS AND METHODS

2.1. Collection of Seaweeds

Padina pavonia, Padina tetrastromatica and Stoechospermum marginatum were collected during low tide, at Hare Island, Thoothukudi from November 2012 to February 2013. The samples were washed thoroughly with seawater followed by fresh water to remove the sand particles and macroscopic epiphytes. After draining, the seaweeds were shade – dried, powered, sieved and used for the preparation of seaweed concentrate.

2.2. Preparation of Seaweed Liquid Fertilizer for Soil Application

About 20g dried seaweed powder with 200ml distilled water was added. It was heated to 60° C and maintained at the temperature for 24h using a hot air oven. The extract was filtered and then centrifuged at 5000 – 10000 rpm to remove most of the suspended impurities. The filtrate was stored in air tight bottles at 4°c (100% seaweed concentrate) [12]. Suitable dilution (1%) was used for soil application on leafy green vegetable crop, *Amaranthus caudatus L*.

2.3. Experimental Design

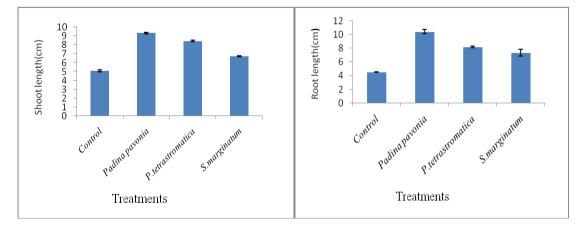
A pot culture experiment was conducted during February to April 2013 at Plant Research Centre, St. Mary's College Campus, Thoothukudi. The pots were filled with 3kg of garden soil. 50 seeds were sown in each pot. After the emergence of seedlings, they were thinned to ten plants per pot and allowed to grow for a period of 30 days. Weeding and watering were done at regular intervals throughout the experimental period. SLF was applied in soil (along with 100ml of distilled water in the ratio of 1: 10) after expansion of first leaf and was continued for twenty days.

2.4. Analyses

Chemicals of analytical grade were used for all the analyses. Plants from each treatment were randomly drawn to study the growth parameters, photosynthetic pigments, biochemicals and antioxidant activities. Growth contributing parameters such as shoot and root length, number of leaves/plant, leaf area and above ground biomass were measured. Root length and shoot length were measured using ruler and recorded in centimeter. Leaf area was measured by using graph paper method as described by [13]. Total chlorophyll and carotenoid [14], total soluble protein [15], carbohydrate [16], free amino acid [17], vitamin C [18], total phenolic content [19], flavonoids [20], DPPH scavenging activity [21], super oxide radical scavenging activity [22] and ferric ion reducing power assay [23] were also recorded. Leaves harvested from 30 days old plants were used for all analyses.

2.5. Statistical Analysis

Data collected in this study was analysed by using Microsoft Excel 2007. One way ANOVA was used to compare differences in the means of total chlorophyll, carotenoid, carbohydrate, free amino acid, soluble protein, vitamin C, total phenolic content, flavonoids, DPPH scavenging activity, superoxide radical scavenging activity and ferric ion reducing power assay of control and treated plants. A significant difference was considered at the level of p<0.05.



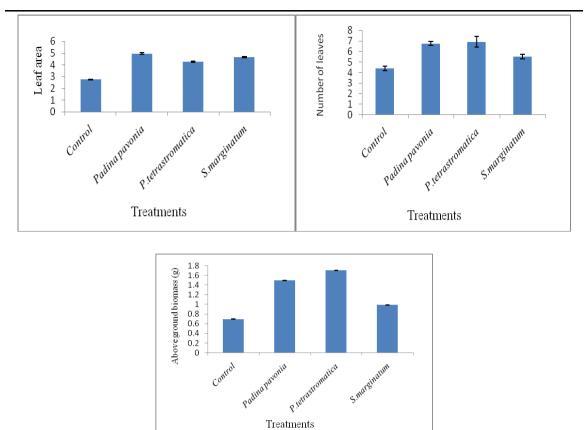


Fig1. SLF (1%) was applied for 30 days. Each value represent the mean of 10 replicates \pm SD.

Table1. Effect of soil application of SLF on photosynthetic pigments, phytochemical and biochemical constituents of Amaranthus caudatus L.

Soil application						
Parameters	Control	Padina pavonia	Padina	Stoechospermum		
			tetrastromatica	marginatum		
Carotenoids (mg/g FW)	41.71±0.6	60.73±0.4*	54.24±0.7*	66.96±0.5*		
Total phenolic content (mg	2.18±0.04	3.03±0.02*	3.10±0.004*	3.31±0.06*		
GAEs/g FW)						
Flavonoid (mg QEs/g FW)	35.56±1.3	60.47±1.3*	68.22±1.5*	57.98±0.8*		
Vitamin C(mg AAEs/g	3.84±0.10	5.11±0.08*	5.36±0.1*	5.54±0.9*		
FW)						
Total soluble protein (mg/g	1.9±0.05	2.3±0.02*	2.8±0.03*	3.1±0.007*		
FW)						
Total carbohydrates	26.99±0.5	39.15±0.7*	43.62±0.4*	42.12±0.3*		
(mg/g FW)						
Free amino acid (mg/g	4.00±0.03	5.26±0.05*	5.07±0.07*	5.96±0.07*		
FW)						
Total chlorophyll (mg/g	0.80 ± 0.01	1.13±0.006*	1.09±0.003*	1.34±0.008*		
FW)						

*Note: *indicates mean values significant at p<0.05 level.*

Values are the mean of three replicates $\pm S.D$.

All the variables were recorded on 30 days old plants

Control = Control plants were irrigated with water.

SLF = 1% seaweed liquid fertilizer.

mg GAEs/g FW = milligram gallic acid equivalents.

mg QEs/g FW = milligram quercetin acid equivalents.

mg AAEs/g FW = milligram ascorbic acid equivalents.

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Soil application						
Parameters	Control	Padina pavonia	Padina tetrastromatica	Stoechospermum marginatum		
Free radical scavenging activity(%)	37.33±0.6	61.00±1*(63%)	62.66±0.6* (67.8%)	56±0.5*(50%)		
Ferric ion reducing antioxidant activity (Absorbance)	0.2±0.002	0.53±0.002* (165%)	0.48±0.005*(140%)	0.55±0.01*(175%)		
Superoxide radical scavenging activity (%)	44.66±1.5	73±1.2* (63%)	75.33±1* (68%)	69.33±0.8* (55%)		
Standard – Ascorbic acid (1mg/ml) Free radical scavenging activity (DPPH) – 98.27% Ferric ion reducing antioxidant activity (FRAP)- 0.853 Superoxide radical scavenging activity – 65%						

Table2. Effect of soil application of SLF on antioxidant activity of Amaranthus caudatus L.

Note: * *indicates mean values significant at p<0.05 level.*

Values are the mean of three replicates $\pm S.D$.

All the variables were recorded on 30 days old plants.

Values within parenthesis indicate percentage increase over control.

Control = *Control plants were irrigated with water*.

SLF = 1% *seaweed liquid fertilizer.*

3. RESULTS AND DISCUSSION

The results showed that soil application of SLF of among three seaweeds of Padina pavonia increased the shoot length (83%), root length (131%) and leaf area (79%) when compared to control plants. SLF of Padina tetrastromatica enhanced the number of leaves (56.8%) and above ground biomass (145%) with respect to control (Fig.1). Plants can take up biologically active compounds of seaweed concentrates through root and leaves [24 and 25]. The rooting response was attributed to endogenous indoles which were probably present in the aqueous seaweed extract, resulting in healthy root and shoot. [26] Reported that lower concentration of SLF from Stoechospermum marginatum promoted the growth of brinjal and [27] also reported similar effect in Cow pea and [28] reported linear growth of both root and shoot in black gram seeds. The seaweed extracts improve the absorption of nutrients through the roots causing additional and strong overall growth of the plant [29]. In the present investigation, more carotenoid level was recorded in SLF of Stoechospermum marginatum (60%), Padina pavonia (43%) and Padina tetrastromatica (30%) when compared to control plants (Table-1). A similar observation was made in Vigna catajung and Dolichos biflorus [30 and 31], Cyamopsis tetragonoloba [32]. In the present investigation, SLF of Stoechospermum marginatum enhanced the total soluble protein (63%) and free amino acid (49%) when compared to untreated plants (Table 1). It was reported that total soluble protein and free amino acid contents were increased in Vigna mungo [33]. Cyamopsis tetragonoloba [34] and Brassica nigra [35] by SLF of Stoechospermum marginatum and Padina tetrastromatica. In the present study the highest carbohydrate was observed in the soil application of SLF of Padina tetrastromatica (61.6%), Stoechospermum (56.05%) and Padina pavonia (49.9%) than control (Table 1). Total carbohydrate increased upto 50% concentration of SLF of Hypnea musiformis with NPK application in black gram [36], Vigna catajung and Dolichous biflorus [30 and 31]. In the present study soil application of SLF of Stoechospermum marginatum, enhanced remarkably in vitamin C (44.2%), total phenolic content (51.8%) and ferric ion reducing power activity (140%) in relation to control (Table 1 and 2). Vitamin C, one of the most important of all vitamins, plays an important role in various biochemical processes, such as collagen formation, iron absorption and is involved in neurotransmission and immune response in humans [37]. In the present study soil application of SLF of Padina tetrastromatica highly increased flavonoid content, free radical scavenging activity (DPPH) and superoxide radical scavenging activity by 91.8%, 66% and 67% respectively

with respect to control (Table 2). Free radicals are responsible for aging and causing various human diseases. Reactive oxygen species such as hydroxyl superoxide and peroxyl radicals are formed in human cells by endogenous factors and result in extensive oxidative damage which can lead to select degenerate conditions and other human diseases [38]. Phenolic compounds can act as antioxidants by chelating metal ions, preventing radical formation and improving antioxidant endogenous system [39]. Phenols are present in seaweeds and plants are found to be new natural and they are acting as new natural source of antioxidants [40].

4. CONCLUSION

The study revealed that SLF of *Stoechospermum marginatum*, *Padina tetrastrastromatica* and *Padina pavonia* could be effectively used in agriculture as an environmentally healthy organic to increase yield and nutrient status of crop plants.

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REFERENCES

- [1] Camargo J.A. and Alonso A., Ecological and toxicological effects of inorganic nitrogen pollution in aquatic ecosystems: a global assessment. Environment International, 32: 831-849 (2006).
- [2] Metting B., Zimmermann W.J., Crouch I. and Van Standen J., Agronomic uses of seaweed and microalgae. In: Akatsuica, I. (ed.): Introduction to Applied Phycology, pp. 589-627, SPB Academic Press, The Hague, the Netherlands (1990).
- [3] Kaliaperumal N., Marine plants of Mandapam coast and their uses. Regl. Cent. Centrl. Mar. Fish. Res. Ins. Special issue: 40-43(2000).
- [4] Booth E., The manorial value of seaweeds. Bot. Mar. 8: 138 143 (1965).
- [5] Crouch I.J. and Van Staden., Evidence for the presence of plant growth regulators in commercial seaweed products, Plant Growth Regul., 13:21-29 (1993).
- [6] Immanuel R. and Subramanian S.K., Effect of fresh extract and seaweed liquid fertilizers on some cereals and millets. Seaweed Res. Utiln. 21: 91-94 (1999).
- [7] Anantharaj M. and Venkatesalu V., Studies on the effect of seaweed extracts on Dolichos biflorus. Seaweed Res. Utiln. 24: 129 137 (2002).
- [8] Jayaraman J., Jeff N. and P. Zamir P., Commercial extract from the brown seaweed Ascophyllum nodosum reduces fungal diseases in green house cucumber. J. Appl. Phycol.23: 353-361 (2010).
- [9] Khan W., Rayirath U.P. and Subramanian S., Seaweed extracts as biostimulants of plant growth and development. J Plant Growth Regul 28: 386-399 (2009).
- [10] Zhang X. and Ervin E.X., Cytokinin containing seaweed and humic acid extracts associated with creeping bentgrass leaf cytokinins and drought resistance. Crop Sci 44: 1737-1745 (2004).
- [11] Craigie J.S., Seaweed extract stimuli in plant science and agriculture. J. Appl Phycol 23: 371-393 (2011).
- [12] Rama Rao K., Preparation of liquid seaweed fertilizer from Sargassum, Seaweed Research and Utilization Association Workshop on algal products and Seminar on Phaeophyceae in India, Madras, India, 4-7 June (1990).
- [13] Santra, S.C., Chatterjee T.P. and Das (Eds.) A.P., College Botany practical, Arunabha Sen Publication, Calcutta, Vol. 1, pp. 69-70 (1989).
- [14] Arnon D.I., Copper enzymes in isolated chloroplast, polyphenol oxidase in Beta vulgaris. Plant Physiol. 24: 1-15 (1949).
- [15] Lowry O.H., Rosebrough H.J., Fair A.L. and Randall R.J., Protein measurement with the follin-phenol reagent, J. Bio-chem., 193-265-275 (1951).

- [16] Dubois M.K., Gilles J.K., Robers P.A. and Smith F., Colorimetric method for determination of sugar and related substances analytical chemistry, 28:350 (1951).
- [17] Moore S. and Stein W.H., Photometric method for use in the chromatography amino acids, J. Biol. Chem. 176: 367-388 (1948).
- [18] Baker H. and Frank O., Clinical vitaminology methods and interpretations. Wiley, New York. NY, USA. Pp.198 (1968).
- [19] Duan X.J., Zhang W.W., Li X.M. and Wang B.G., Evaluation of antioxidant property of extract and fractions obtained from a red alga. Polysiponia urceolata. Food Chem, 95: 37-43 (2006).
- [20] Zhishen J., Mengcheng T. and Jianming W., The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals, Food Chem, 64: 555-559 (1999).
- [21] Hatano T., Kagawa H., Yasuhara T. and Okuda T., Two new flavonoids and other constituents in licorice root; their relative astringency and radical scavenging effects. Chem. Pharm. Bull 36: 1090-1097 (1988).
- [22] Dasgupta N. and De., Antioxidant activity of Piper betel L. Leaf extract in vitro. Food Chem. 88: 219-214 (2004).
- [23] Oyaizu M., Studies on products of browning reactions; antioxidative activities of products of browning reaction prepared from glucosamine. Japanese Journal of Nutrition 44, 307-315 (1986).
- [24] Nelson W.R. and Van Staden J., The effect of seaweed concentrates on the growth of nutrient-stressed, greenhouse cucumbers, Journals of Horticultural Science 19:81-82 (1984).
- [25] Nelson W.R. and Van Staden J., Effect of seaweed concentrate on growth of wheat. S. Afr. J. Sci. 82:199-200 (1986).
- [26] Vijayanand N., Ashok, V., Rathinavel, S. National Symposium and Exposition on Seaweeds, Cochin. January. Abstract No.57, (2004).
- [27] Sivasankari S., Chandrasekaran M., Kannathasan K. and Venkatesalu V., Effect of seaweed extract on growth and yield of cowpea. Seaweed Res. Utiln. 28: 145-150 (2006).
- [28] Ramamoorthy K. and Sujatha K., Effect of seaweed extract on the ageing of black gram seeds. Seaweed Res. Utiln. 29 : 119-123 (2007).
- [29] Crouch I.J., Beckett R.P. and Van Staden J., Effect of seaweed concentrates on the growth and mineral nutrition of nutrient stress lettuce. Journal of Applied Phycology 2: 269-272 (1990).
- [30] Anantharaj M. and Venkatesalu V., Effect of seaweed liquid fertilizer on Vigna catajung. Seaweed Res. Utiln. 23: 33-39 (2001).
- [31] Anantharaj M. and Venkatesalu V., Studies on the effect of seaweed extracts on Dolichos biflorus. Seaweed Res. Utiln. 24: 129-137 (2002).
- [32] Thirumal Thangam R.T. and Rani S.M.V., Effect of seaweed liquid fertilizer on the growth and biochemical constituents of Cyamopsis tetragonoloba L. Seaweed Res. Utiln. 25: 99-103 (2003).
- [33] Venkatesalu V., and Kalaivanan C., Utilization of seaweed Sargassum myriocystum extracts as a stimulant of seedlings of Vigna mungo (L.) Hepper. Spanish Journal of Agricultural Research 10(2), 466-470 (2012).
- [34] Sivasangari Ramya S., Vijayanand N. and Rathinavel S., Influence of seaweed liquid fertilizer on growth, biochemical and yield parameters of cluster bean plant. J. Green Bioenergy 1(1): 19-32 (2012).
- [35] Kalidass C., Jayarani S. and Glory M., Effect of seaweed liquid fertilizers on growth and biochemical constituents of Brassica nigra (L.). Int J Agric Environ Biol., 3: 307-311 (2010).
- [36] Tamilselvan C, and Kannan L., Studies on the utilization of seaweed as fertilizer for black gram, Indian J. Argic. Res., 28 (2), 121-126 (1994).
- [37] Peng X., Yu. Y. and Tang C., Occurrence of steroid estrogens, endocrine-disrupting phenols and acid pharmaceutical residues in urban river water of the Pearl River Delta, South China. Sci Total Environ., 397, 158-166 (2008).

- [38] Abu-Ghannam N. and Gupta S., 'An assessment of the antioxidant and antimicrobial activity of six species of edible Irish seaweeds.' Int. Food Res. J., 17, 205-220 (2010).
- [39] Al-Azzawie H.F. and Alhamdani M.S. Hypoglycemic and antioxidant effect of oleuropein in alloxan-diabetic rabbits. Life Sci 78:1371–1377 (2006).
- [40] Duan, S.W., Bianchi, T.S. and Shiller, A.M., Seasonal changes in abundance and composition of dissolved and particulate organic matter in the lower Mississippi and Pearl Rivers. Abstracts of Papers of the American Chemical Society, vol. 225. U925-U925 087-GEOC Part 1 (2003).

AUTHORS' BIOGRAPHY

Mrs.T. Kumareswari, M.Sc.,B.Ed., PGDCA and M.Phil., in Botany, doing Ph.D under the guidance of Dr. S. Maria Victorial Rani, Associate professor, HOD and Dean of Research, St. Mary's College(Autonomous), Thoothukudi. She is a member of Seaweed Research and Utilization Association (SRUA). She has participated 8 seminar/conference/workshop in national and international level and presented 2 research papers in reputed national and international journals. She is doing research on utilization of seaweeds as a biofertilizer.



Dr. S. Maria Victorial Rani, Associate Professor and Head, Department of Botany, working in St.Mary's College (Autonomous), Thoothukudi, Tamil Nadu, India, has 33 years of teaching and 29 years of Research in environmental physiology and seaweeds. She has published 45 research articles in reputed national and international journals and presented more than 20 research papers in national and international seminars and conferences. She was the member in

Faculty selection committee, FDP programme XI plan. She served as examiner question paper setter for UG and PG Botany programme at University and College level. She has completed a Major Research Project in 2014 funded by UGC, New Delhi. Under her headship the department has organized a two day exhibition on herbal plants, for Higher secondary School children in 2014. At present she is the member of Senate Manonmaniam Sundaranar University, Tirunelveli, Dean of Research St. Mary's College (Autonomous), Thoothukudi and holds many positions in administration too.