1. INTRODUCTION

Indonesia always imports soybean annually to meet a demand for food industries, namely tempe, tofu and soy sauce. For example, Indonesia imported 2.26 million ton soybean in 2015 which is equivalent to US$ 1.03 million (Kompas 2016). A significant decrease of national soybean production is caused by unfavorable price of soybean seed. Under such undesirable condition for growing soybean, farmers shift to grow other crops, such as corn and horticultural crops. An increase of soybean price to meet the benefit of the farmers and consumers would drive the farmers’ motivation to grow soybean in a larger area. In such circumstances, it is expected to increase the National soybean production.

Japanese government subsidizes soybean price to farmers as much as 11,310 yen/60 kg soybean (Yusman 2017) in order to motivate farmers growing soybean. However, this strategy seems to be unlikely to happen in Indonesia. It seems to be possible to increase the domestic soybean price through better awareness and understanding of the Indonesian people that soybean seeds generated by a conventional breeding is safer and better to be consumed. In addition to high protein content, soybean produces secondary metabolites playing a role as antioxidant (Wu et al. 2009; Doss et al. 2011; Mukić et al. 2011; Sefatie et al. 2013; Pertiwi and Nurhidayah 2013). An intake of soybean would likely to improve antioxidant state in human body (Bakhtiari et al. 2019). The ample amount of antioxidant in human body is required to negate the excessive amount of reactive oxygen which is harmful to human health. Our conventionally bred soybean, yellow seed coat, was reported to possess significantly higher antioxidant activity than imported soybean (Soedarjo et al. 2019). These reasonings should be disseminated to the people to acquire a special appreciation. Such a perception could hopefully trigger the people to purchase soybean seeds obtained from conventional breeding with relatively higher price benefitting to soybean growers. As a result, farmers will grow soybean on much larger area and the National soybean production will significantly increase, consequently.

Other than yellow soybean seeds, Indonesian Legume and Tuber Crops Research Institute (ILeTRI), has released some black soybean varieties, Detam 1, Detam 2, Detam 3 and Detam 4 (ILeTRI 2016). Researchers (Takahashi et al. 2005; Tilić et al. 2011; Malencic et al. 2012; Dajanta et al. 2013;
Yusnawan (2016) have revealed higher antioxidant activity of black soybean seed than yellow soybean seed. Indeed, our black soybean seeds have also been assayed to show higher antioxidant activities than the yellow soybean varieties (Yusnawan 2016). However, it has not been investigated where the antioxidant activity is mainly concentrated, in the seed coat or in the cotyledon of the soybean seed. The present research work was carried out to evaluate the phenolic content and the degree of antioxidant activities on the seed coat and the cotyledon of black soybean seeds.

2. MATERIALS AND METHOD

The present laboratory research work was done at the Chemistry laboratory, the Institute of 10 November Surabaya, East Java, Indonesia in 2018-2019. The improved black soybean seeds used in the present work were provided by the Germplasm Division of ILeTRI, The Indonesian Ministry of Agriculture (Table 1 and Figure 1). Beside the whole grain, Detam 1 and Detam 2 were partitioned into seed coat and cotyledon (seed without the seed coat) (Figure 4). Each sample of soybean seed was macerated into powder and extracted with methanol before further use for phenolic and antioxidant evaluation.

Table 1. Indonesian improved soybean varieties used in the present research work

<table>
<thead>
<tr>
<th>No.</th>
<th>Varieties</th>
<th>Genetic background/breeding</th>
<th>Seed coat color</th>
<th>Seed size*</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Detam 1</td>
<td>Introduction line x Kawi</td>
<td>Black</td>
<td>Big</td>
</tr>
<tr>
<td>2</td>
<td>Detam 2</td>
<td>line 9837 x Wilis</td>
<td>Black</td>
<td>Medium</td>
</tr>
<tr>
<td>3</td>
<td>Detam 3</td>
<td>W 9837 x Cikurai</td>
<td>Black</td>
<td>Medium</td>
</tr>
<tr>
<td>4</td>
<td>Detam 4</td>
<td>W 9837 x G100H</td>
<td>Black</td>
<td>Medium</td>
</tr>
<tr>
<td>5</td>
<td>Mallika</td>
<td>Selected from local variety, Central Java, Indonesia</td>
<td>Black</td>
<td>Medium</td>
</tr>
</tbody>
</table>

* Big: ≥ 14 g/100 seeds, Medium: 10-14 g/100 seeds

2.1. Determination of Total Phenolic Compound

The powder of each soybean samples was dissolved into methanol to reach a final concentration of 1 mg/ml and was considered as a sample. Sixty six microliter of each sample and 500 μl reagent of 10 % Folin was mixed thoroughly. Afterwards, the mixture was incubated in the dark room for 5 minutes. After incubation, the mixture was added with 500μl of 6% Na2CO3 and was stirred thoroughly. The mixture was further incubated in the dark room for 90 minutes. Finally, each sample was determined by spectrophotometer at λ 750 nm. Measurement of each sample was done 3 times and was presented as mg GAE/g sample (Qassabi et al. 2018).

2.2. Determination of Antioxidant Activity by ABTS

Free radical solution of ABTS (2,2’-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) was prepared by mixing 5 ml of 7 mM ABTS and 88 μl of 140 mM K2S2O8 (Putri et al. 2018). This Free radical solution was incubated in the dark room at room temperature for 16 hours. Afterwards, 300 ml ethanol was added into the free radical solution in order to reach absorbance of 0.7 ± 0.02 unit at λ 734 nm. The sample solution consisting of 10 μl sample (10 mg sample/ml DMSO) and 1 ml of ABTS free radical solution was incubated for four minutes at room temperature. Furthermore, the sample solution (As) was measured by UV-Vis spectrophotometer at λ 734 nm. Trolox (6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid) was used as a positive control. DMSO was used as a negative control or blank (Ab). Each sample was measured three times and standard deviation was calculated based on the three replicates. The inhibitory activity was measured by formula (1).

Inhibitory activity (%) = \([(Ab – As)/Ab]\) x 100%  (1)
2.3. Determination of Antioxidant Activity by DPPH

Antioxidant activity was measured by employing the method of Brand Williams et al. (1995) after slightly modification by Dudonne et al. (2009). The methanolic extracts of soybean seeds was dissolved in methanol (analytical grade) to maximum concentration. A DPPH solution (2,2-diphenyl-2-picrylhydrazyl hydrate) was dissolved in methanol to a final concentration of 6 x 10^{-5} M. Extract of each sample (33.3 ml) was poured into a test tube containing 1 ml DPPH, the mixture was stirred thoroughly and was named as ‘As’. The negative control consists of 33.3 ml methanol and was thoroughly mixed with 1 ml DPPH in a test tube and this mixture was named as ‘Ab’. Gallic acid was used as positive control. The mixture of all samples (As) and the mixture of the negative control (Ab) were incubated at room temperature for 20 minutes. Right after the incubation, the absorbance of the mixture of each sample (As) and the mixture of Ab were read with a spectrophotometer at 515 nm. The reading was done at three replicates for each sample and standard deviation was calculated based on the three replicates. The degree of antioxidant activity was calculated as formula 1.

3. Results and Discussion

3.1. Total Phenolic Content

Total phenolic content among Indonesian black soybean seeds varied significantly (Figure 2). The highest total phenolic content was observed on the seed of soybean var. Detam 2 (25.94 g GAE/g). Soybean var. Detam 1 contains the lowest total phenolic content, i.e., 4.51 mg GAE/g which is significantly lower compared to phenolic content of Detam 2, Detam 3, Detam 4 and Mallica. Whilst, the phenolic contents of Detam 3, Detam 4 and Mallica were comparable. Black soybean seed has been reported to be related to relatively high phenolic compound and was confirmed to contain higher phenolic content than yellow soybean seed coat (Takahashi et al. 2005; Kumara et al. 2010; Ţilić et al. 2011; Malencic et al. 2012; Dajanta et al. 2013; Yusnawan 2016). However, our present results indicate that the black soybean seed does not always possess high phenolic content. Phenolic content of Detam 1 (black soybean seed) was only half of the phenolic content of Indonesian yellow soybean seed, Detap 1 and Demas (Soedarjo et al. 2019). Therefore, the genetics encoding for phenotypes other than the color of the soybean seed coat might also contribute to the phenolic profile.

3.2. Antioxidant Activity by ABTS and DPPH Methods

Figure 3 depicted a variability of antioxidant activities among the black soybean seeds tested as measured by ABTS as well as by DPPH methods. Soybean seed of Detam 2 showed the highest antioxidant activity as measured either with ABTS or DPPH. In contrary, Detam 1 revealed the lowest antioxidant activity, eventhough, the seed coat color is also black. All soybean varieties used in the present research work were generated through a conventional breeding with different genetic backgrounds (see Table 1). The previous findings revealed that different genetic background within
In Vitro Investigation on Phenolic Compound and Antioxidant Activities from Methanolic Extracts of Black Soybean Seed

Figure 3. Aktivitas antioksidan biji kedelai warna kulit hitam dengan metode ABTS (99 μg/ml) dan DPPH (619 μg/ml)

plant species caused a variability of antioxidant measured with ABTS (Mikulajová et al. 2007; Sing et al. 2018). A variability of antioxidant activity within the black soybean varieties was also reported due to different genetic background (Kumara et al. 2010; Lee et al. 2016, Yusnawan 2016). Considering these previous findings and the results of the present research work, it is suggested that genetic background determining the phenotypes other than the genetics for seed coat color seems to play role in the degree of antioxidant activity.

3.3. Correlation Between Phenolic Compound and Antioxidant Activity

Correlation value (r) between total phenolic compound and antioxidant activity was found to be highly correlated as measured with ABTS (r = 0.922104459) and DPPH (r = 0.961998886) methods and is significant at 1% (Table 2). The result of this correlation analysis suggests that high phenol content of soybean seed will be followed with high antioxidant activity measured with either ABTS or DPPH method. The Present study is in accordance to the research results reported by several researchers who found a positive correlation between total phenol and antioxidant activities in soybean and other plant species (Malenčić et al. 2007; Mikulajová et al. 2007; Stanković 2011; Lee et al. 2014; Nam et al. 2014; Rebaya et al. 2014; Hossain and Shah 2015; Fidrianny et al. 2016; Khalid et al. 2017; Parikh and Patel 2017; Johari and Khong 2019). Therefore, it could be suggested that total phenol is a good initial indicator of antioxidant activities and could be used as an important trait to screen soybean genotypes exerting certain degree of antioxidant activity.

Table 2. Correlation analysis between phenolic compound and antioxidant activities within the black soybean seeds.

<table>
<thead>
<tr>
<th></th>
<th>Antioxidant by ABTS</th>
<th>Antioxidant by DPPH</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total phenol</td>
<td>r = 0.922104459**</td>
<td>r = 0.961998886**</td>
</tr>
<tr>
<td>Note</td>
<td>Highly correlated and significant at F 1%</td>
<td>Highly correlated and significant at F 1%</td>
</tr>
</tbody>
</table>

3.4. Localization of Antioxidant Activities within Soybean Seeds

The color of cotyledons (seed without seed coat) of all black soybean seed is yellow (figure 4). Yellow soybeans were found to reveal lower antioxidant activity than black soybean (Kumara et al. 2010; Lee et al. 2016, Yusnawan 2016). Therefore, it is of the interest of the present research work to investigate if the black seed coat contributes to relatively higher antioxidant activity. The seeds of Detam 1 and Detam 2 (the black soybean seeds) were partitioned into the whole seed (H), the cotyledon or seed without the seed coat (S) and the seed coat (SC). Each part of the seed was assayed for antioxidant activity measured with DPPH. The result of the present research work depicted the highest antioxidant activity on the seed coat of both Detam 1 and Detam 2 (Figure 5). The seed coat (SC) of Detam 1 and Detam 2 exerted approximately 7.6 times and 6 times antioxidant activity than their cotyledons (S), respectively. It is a new evident that high antioxidant activity of the black soybean was due to the seed coat. For making Tempe and Tofu, the seed coat of soybean is usually discarded. Considering the antioxidant is mainly localized within the seed coat of black soybean seed, therefore, the seed coat of black soybean could be used as a good source of antioxidant.
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Figure 4. The whole seed (H) and the seed without seed coat, cotyledon (SC) of Detam 1 and Detam 2

Figure 5. Antioxidant activities of black coated soybean seeds as measured with DPPH (319.5 μg/mL).

H, S and SC are whole seed, seed without seed coat (cotyledon) and seed coat.

4. CONCLUSION

Methanolic extracts of Indonesian black soybean seeds contained various amount of phenolic content and showed different antioxidant activities. Black soybean seed did not always show comparably high antioxidant activities. Antioxidant activities positively and highly correlated to the phenolic content. High phenolic content of soybean seed is followed with high antioxidant activity. Thus, phenolic content could be used as an initial indicator to distinguish the degree of antioxidant activities. In black soybean seed, high antioxidant was mainly located within the seed coat. Soybean seed coat could be used as a better source of antioxidant.

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