

Hamdy .F.M. El-shamy¹, Y. A. Aggour¹, A. M. Ahmed²

¹Department of Chemistry, Faculty of Science, Damietta University, Egypt

²Department of Chemistry, Faculty of Science, Alexandria University, Egypt

*Corresponding Author: Hamdy. F.M El-shamy, Department of Chemistry, Faculty of Science, Damietta University, Egypt

Abstract: Carbon steel has wide applications in industry (especially in petroleum industry, eg. pipelines out from the top of distillation towers (over head pipes), pressure vessels, nozzles etc...) and domestic life. Study the phenomenon of electropolishing which is important as metallic finishing process which takes place before electroplating of metal. This work is devoted to study the electropolishing of carbon steel in phosphoric acid in presence of polymers molecules (Starch, Gelatin, Agar, Chitosan, Arabic gum, Cellulose acetate, Cellulose and Carboxymethyl cellulose sodium salt). The rate of electropolishing of carbon steel in phosphoric acid solution in case of applied various affecting factors was investigated by estimated polarization curves from values of current and voltage that were abtained from the electrochemical cell. Morphology of the specimens after the experiment is monitored using scanning electron microscope (SEM). The (SEM) micrographs support the experimental measurements and give more light on carbon steel surface. Finally, samples of industrial waste water are collected from a drain of vaccum distillation tower over head pipes in Alexandria petroleum company and ferrous ions Fe+2 concenterations are determined in case of presence and absence of polymers molecules in order to judge the role of these polymers as a corrosion inhibitors on the ability to control the corrosion on carbon steel over head pipes.

Keywords: *Metallic finishing process, polymers molecules, Electropolishing of carbon steel, scanning electron microscope, corrosion inhibitors*

1. INTRODUCTION

Electropolishing is suitable for polishing of hardened materials that are mechanically difficult to machine, because in electro polishing the electrode and the work piece are not in contact with each other. The mechanism of electro polishing is usually explained as follows: the electro polishing effect occurs because of differential dissolution, as the current is applied, the oxidization film covering the lower peaks of the surface, which has a high specific [1].

In view of its strength and low cost, carbon steel is used widely as a material of construction in chemical industry to build pipelines, storage tanks, heat exchangers, pumps, etc [2]. However steel suffers from corrosion in electrolytes because of the presence of cementite in its structure [3,4].

Cementite $[Fe_3C]$ which does not form solid solution with iron [Fe] and hence leads to steel corrosion through the formation of a multitude of microscopic galvanic cells of the type : $Fe / Electrolyte / Fe_3C$ (Cementite)

To protect steel structures against corrosion different techniques are used such as cathodic protection, anodic protection, coating and the use of inhibitors **[4-7]**.

Inhibitors are materials that may be injected into the system. They plate out on the surface and inhibit the formation of corrosion cells. They are commonly used in pipelines and other vessels that will contain materials that are corrosive [8,9]. Some metals, such as gold and platinum, corrode very slowly or not at all. Choosing a corrosion resistant material can reduce the rate of corrosion. They are sometimes injected into the water stream that may be used for the surface preparation of steel, as in the case of water jetting.

In the formulation of some primers inhibitive pigments are used. These inhibitive pigments inhibit, or interfere, with the corrosion process.

To improve the protective efficiency of carbon steel corrosion in corrosive environment, great efforts have been put into the investigation and lots of technologies have been used. The inhibitive power of the organic inhibitors has been interpreted in term of many different characteristics such as molecular size, molecular weight, molecular structure, nature of heteroatom present in the molecule, their affinities to get adsorbed on the metal surface, etc [10].

A good number of the efficient corrosion inhibitors are organic compounds that contain nitrogen, oxygen, sulfur, phosphorus, and multiple bonds or aromatic rings in their structures [11]. The molecular electronic structures and electron densities around these functional groups are the key structural features that determine the effectiveness of inhibition (i.e. specific interaction between functional groups and metal surface and heteroatom like N, O and S play an important role in inhibition due to the free electron pairs which they possess). When both of these features combine, increased inhibition can be observed [12].

Polymers as accrossion inhibitors were studied in this work. The inhibiting action of these natural polymers is usually attributed to their interaction with carbon steel surface via their adsorption. Polar functional groups are regarded as the reaction center that stabilizes the adsorption process. In general, the adsorption of inhibitor on metal surface depends on the nature and the surface charge of the metal, the adsorption mode, its chemical structure and the type of electrolyte solution [13]. The aim of the present work is to study the effect of natural polymers namely starch, gelatin, agar, chitosan, cellulose acetate, cellulose and carboxymethyl cellulose sodium salt on the rate of electropolishing of carbon steel

A great deal of work has been directed to study the mechanism of electropolishing as well as establishing the optimum polishing conditions. Studies have revealed that electropolishing is a diffusion controlled process and its efficiency is determined by rate factors. Knowledge of the reaction kinetics is essential for the most economic utilization of the process.

The rate of electropolishing of carbon steel in phosphoric acid solution in absence and presence of polymers molecules was studied in this article by measuring the limiting current of electropolishing under natural convection condition as a function of electrode height, phosphoric acid concentrations, different concentrations of polymers molecules and temperature, in order to indicate the role of polymers molecules on carbon steel electropolishing and to correlate the activation parameters. Dimensional analysis were investigated to correlate mass transfer during electropolishing reaction in presence of polymers under forced convection condition (by using rotating cylinder electrode RCE). Physical properties of solution such as density and viscosity were studied [14].

2. EXPERIMENTAL PROCEDURE

2.1. Chemicals

Five different H_3PO_4 concentrations were used, namely 4, 6, 8, 10, and 12 M, all solutions were prepared using A.R grade chemical. Blank solution (8M H_3PO_4) was prepared as well as in presence of different concentrations from the polymers in the range of concentration (100-800 ppm). The limiting current was determined at four different temperatures 298, 303, 308 and 313 K. The polymers selected in the present work are Starch, Gelatin, Agar, Chitosan, Arabic gum, Cellulose acetate, Cellulose and Carboxymethyl cellulose sodium salt (CMC). Supplied by SIGMA-ALDRICH Chemicals Ltd.

The choice of the polymers molecules is based on the following considerations:

- Natural products
- Non-toxic compounds
- Low price
- Easy available
- Environmentally friendly compounds

Analysis of Carbon Steel Specimens Composition (wt %)

Components	wt %
С	0.2
S	0.04
Mn	2.6
Р	0.039
Si	0.36
Fe	96.761
Total	100

2.2. Measurements of the Limiting Current

The limiting current was determined by polarization curves were plotted by increasing the cell current stepwise and measuring the corresponding steady state anode potential **[14,15]**. Anode potential was measured against a reference steel electrode placed in the cup of luggin tube whose tip is placed 0.5 mm from the anode surface. The potential difference between the anode and the reference electrode was measured by voltmeter. The temperatures were regulated by placing the cell in a constant $\pm 0.05^{\circ}$ C ultra thermostat.

The rate of electropolishing of carbon steel was determined at 25, 30, 35 and 40° C. Polarization curves were constructed for Starch, Gelatin, Agar, Chitosan, Arabic gum, Cellulose acetate, Cellulose, Carboxymethyl cellulose sodium salt(CMC) - acid mixtures at different concentrations.

2.3. Surface Finish Morphology Measurement

The surface finish after EP was characterized using scanning electron microscopy (SEM). The SEM images were taken using a JEOL, JSM-5300, scanning microscope, OXFORD instrument.

SEM is one of the most widely used analytical tools, due to the extremely detailed images it can provide. Coupled to an auxiliary Energy Dispersive X-ray Spectroscopy (EDS) detector, this technique also offers elemental identification of nearly the entire periodic table. The Scanning Electron Microscope (SEM) is a microscope that uses electrons rather than light to form an image. The SEM has a large depth of field, which allows a large amount of the sample to be in focus at one time. The SEM also produces images of high resolution, which means that closely spaced features can be examined at a high magnification. Preparation of the samples is relatively easy since most SEMs only require the sample to be conductive to not charge and deflect the electron. The combination of higher magnification , larger depth of focus, greater resolution, and ease of sample observation makes the SEM one of the most heavily used instruments in research areas today[16].

2.4. Atomic Absorption Spectroscopy Technique

(Agilent-280FS AA) Atomic Absorption Spectrophotometer were used at 248.3 nm with Iron Hollow-Cathode Lamp to get the concentration of iron in the samples to evaluate these polymers as corrosion inhibitors. The 280FS AA is external computer-controlled atomic absorption spectrometers supporting multielement flame AA determinations using Fast Sequential analysis for improved sample throughput with flame AA. The supplies of fuel (acetylene) and oxidant shall be maintained at pressures somewhat higher than the controlled operating pressure of the instrument by suitable valves [17,18].

3. RESULTS AND DISCUSSION

3.1. Leveling Process

Leveling is the principle process in electro-polishing [17] and can be explained by mass transfer mechanism [18]. The study of leveling is based on the classical current voltage curves of electro dissolution.

Figures1&2 shows a typical polarization curves obtained using different concentration of electrolyte and using different electrode height. We found that the rate of electropolishing, which is represented the rate of steel corrosion in phosphoric acids decreases with increasing electrode height and electrolyte H_3PO_4 concentration.



Figure 1. Polarization curves obtained using different concentration of electrolyte in case of undivided cell.



Figure 2. Polarization curves obtained using different electrode height in case of undivided cell.

3.2. Influence of Temperature and Activation Energy During Electropolishing

the influence of temperature on the value of the limiting current that determines the carbon steel electropolishing rate in absence and presence of different concentrations of the studied polymers in case of undivided and temperature range 298-313 K were studied.

The influence of temperature and activation energy (E_a) of electropolishing process in presence of polymers was calculated using Arrhenius-type plot equation [19,20]

$\mathbf{I}_{t} = \mathbf{A} \exp \left(-\mathbf{E}_{a} / \mathbf{RT}\right)$

where A is a modified frequency factor (pre-exponential factor), E_a is the activation energy (kJ mol⁻¹), R is the gas constant (8.314 J/K mol), and T is the absolute temperature (K). The plot of ln I_ℓ versus reciprocal of absolute temperature, 1/T, gives a straight line with slope equal to $-E_a$ /R as shown in Figure (3) for cellulose acetate as example, from which the activation energy for the electropolishing process can be calculated

We observed that the limiting current increases with increasing temperature for different concentrations of polymers, with a consequent increase in the rate of electropolishing process of carbon steel [21,22].

This increase could be explained due to:

- 1. The decrease of the strength of adsorption processes at elevated temperature and suggested a physical adsorption mode.
- 2. Increasing the temperature of the system could not augment the interaction between the metal surface and the inhibitor leading to lower surface coverage, therefore the inhibiting effect decrease [23].

(1)

3. The viscosity of the solution decreases with increasing temperature with consequence increase in the diffusivity of Fe^{2+} according to equation (3.3) [19,23].

The activation energy of the process is an important parameter for determining the rate controlling step. If adsorption of the species on the reaction surface and subsequent chemical reaction takes place (E_a) values usually > 43 kJ mol⁻¹. If the rate controlling step is the diffusion of the aqueous species in the boundary layer indicating physical adsorption (E_a) is generally \leq 43 kJ mol⁻¹. In this work it is noted that adsorption of the species on the reaction surface in case of undivided cell were take place[24].



Figure3. Arrhenius-relationship between limiting current and temperature for solution contain different polymers at different concentrations in case of undivided cell

3.3. Variation in the Rate Within A Reaction Series

Changes in either, or both, the enthalpy or the entropy of activation may be cause variation in the rate within a reaction series. The correlation of ΔH^* with ΔS^* is a linear relationship may be stated algebraically as [25]:

$\partial \Delta G^* = (1 - T/\beta). \partial \Delta H^*$

If follows that when $\delta \Delta G^*$ equals zero, (β) equals (T). In other words, the slope in a linear plots of ΔH^* against ΔS^* is the temperature at which all reactions that confirm to the line occur at the same rate, (β) is therefore known as the isokinetic temperature represented in Kelvin.

Figure (4) shows that the isokinetic plot of ΔH^* against ΔS^* for polymers was found to be linear and the isokinetic temperatures (β) were computed from the slopes of these plots as shown in Table (1). Which smaller than 298 K indicate that the rate of the reaction is entropy control, (Starch, Agar, Arabic gum) in case of Undivided cell while which is greater than 298 K indicate enthalpy control reaction(Gelatin, Chitosan,Cellulose acetate,Cellulose and CMC in case of Undivided cell & all compounds in case of divided cell) [26].

Table1. Relation between ΔH^* and $-\Delta S^*$ at 8M H_3PO_4 and 3cm height for different concentrations of polymers in case of using (a) undivided cell and (b) divided cell.

Type of	Isokinetic temperatures (K)								
Cell	Starch	Gelatin	Agar	Chitosan	Arabic gum	Cellulose	Cellulose	CMC	
						acetate			
Undivided	238	342	239	334	247	347	338	341	
Cell									
Divided	380	353	367	331	349	338	336	330	
Cell									

(2)



Figure4. Relation between ΔH^* and $-\Delta S^*$ at 8M H₃PO₄ and 3cm height for different concentrations of gelatin in case of using undivided cell

3.4. Adsorption Isotherm

Figure (5) shows the Flory Huggins adsorption isotherm for cellulose acetate plotted as $\log \theta/C$ vs. $\log (1-\theta)$ that are reported as shown in Tables (2) yielded straight line with slope (x) and intercept (xK) for polymers in case of using undivided cell and divided cell at 298 K. The calculated values of (x) and (K) are given in Table (3). The values of (x) depend on the type of polymers. It is clear that the experimental data fits Flory Huggins adsorption isotherm (except Gelatin and Agar in case of using divided cell). The data of Table (3) reveals that, the values of (x) for (Starch, Agar, Chitosan, Arabic gum, Cellulose acetate, Cellulose and CMC) in cae of using undivided cell were higher than one, implied that one inhibitors molecules replace more than one water molecule at constant temperature [26,27]. On other hand the other values of (x) indicated that one inhibitor molecule replace one water molecule onto metal surface.

Figure (6) shows Temkin adsorption isotherm for cellulose acetate plotted as θ vs.log C for polymers. Linear plots obtained indicating that the experimental results at all additives studies obey the Temkin adsorption isotherm. The adsorptiondata deduced from Temkin adsorption isotherm in case of using undivided cell are presented in Tables (2). The molecular interaction parameter "a" can have both positive and negative values indicates attraction forces between absorbed molecules. It is seen in Tables (3) that the values of "a" in all cases are negative indicating that repulsion exists in the adsorption layer. It is generally known that K denotes the strength between the adsorbate and adsorbent. Large values of K imply more efficient adsorption [27].

Figure (7) shows Kinetic adsorption isotherm for cellulose acetate where y is the slope of the linear relation between log ($\theta/1-\theta$) vs. log (C) for all polymers compounds that are reported as shown in Tables (2), if the slope is greater than unity implies the formation of multi-layers of the additive on the metal surface, while if less than unity, however, it means that the given additive molecule occupied more than one active site. Values of y and number of active sites 1/y of the metal surface that occupied by one molecule of the polymer additives under the present conditions have given in Tables (3).

Obviously, it was concluding from the values in Tables (3) that the number of additive molecules which occupy one active site in some cases less than unity and in other cases greater than unity than unity [26,28]. Also the efficiency of a given inhibitor was essentially a function of the magnitude of its binding constant or the equilibrium constant K, large values of K indicate better and stronger interaction, whereas small values of K mean that the interaction between the inhibitor molecules and the metal is weaker. Hence, according to the numerical values of K as in Table (3), it is observed that the higher value of K of Cellulose acetate compared to other polymers. This indicate strong adsorption of Cellulose acetate on anodic carbon steel surface and more stable adsorbed layer is formed which increase the inhibition efficiency of Cellulose acetate relative to other polymers[28,29].

The adsorption of inhibitors at anodic metal surface might be due to the formation of either electrostatic (physisorption) or covalent bonding between the inhibitor and metal surface (chemisorptions) [30], and this depend on the standard free energy of adsorption (ΔG^{o}_{ads}).

International Journal of Advanced Research in Chemical Science (IJARCS)

The standard free energy change for the adsorption process (ΔG^{o}_{ads}) for the different inhibitors can be calculated from the following equation [15,31]:

$\Delta \mathbf{G}^{\mathbf{o}}_{\mathbf{ads}} = -\mathbf{RT} \ln (\mathbf{55.5} \mathbf{K})$

(3)

The values of ΔG^{o}_{ads} in case of using undivided cell and divided cell are collected in Table (4). It is clear from Table (4) that the addition of polymers cause negative values of ΔG^{o}_{ads} , which indicate that the polymers molecules are adsorbed spontaneously onto the carbon steel surface in phosphoric acid and there is strong interaction between inhibitors and carbon steel surface which indicated that all inhibitors strongly adsorbed on carbon steel sueface [32]. It is generally accepted that, the values of ΔG^{o}_{ads} are more positive than - 40 kJ mol⁻¹(i.e. the ΔG^{o}_{ads} values are less than-40 kJ mol⁻¹), meaning that the polymers molecules are physically absorbed through electrostatic interactions between the polymers molecules and the metal surface [33]. The results show that Cellulose acetate, CMC, and Chitosan give maximum efficiency and show more negative free energy of adsorption than other polymers indicating that it is strongly adsorbed on the metal surface [33,34].

It is also noticed from Table (4) that the order of decreasing in the standard free energy of adsorption (ΔG^{o}_{ads}) is as follows:

 $Cellulose \ acetate > (CMC) > Chitosan > Cellulose > Arabic \ gum > Starch > Agar > Gelatin$

Table2. The limiting current and surface coverage in 8M phosphoric acid with different natural polymers concentrations at 298 K

Natural	С	IL	θ	(1-θ)	θ/(1-θ)	log(1-θ)	$\log\theta/(1-\theta)$	log θ/C	log C
polymers	(ppm)	(mA)						_	
additives									
Starch	100	885	0.132	0.868	0.152	-0.061	-0.818	0.121	-1.000
	200	850	0.167	0.833	0.200	-0.079	-0.698	-0.078	-0.699
	300	780	0.235	0.765	0.307	-0.116	-0.512	-0.11	-0.523
	400	700	0.314	0.686	0.458	-0.164	-0.339	-0.105	-0.398
	500	660	0.353	0.647	0.546	-0.189	-0.263	-0.1512	-0.301
	600	600	0.412	0.588	0.701	-0.231	-0.154	-0.1632	-0.222
	700	580	0.431	0.569	0.757	-0.245	-0.121	-0.2106	-0.155
	800	550	0.461	0.539	0.855	-0.268	-0.068	-0.2394	-0.097
Gelatin	100	950	0.069	0.931	0.074	-0.031	-1.130	-0.1611	-1.000
	200	900	0.118	0.882	0.134	-0.054	-0.873	-0.2291	-0.699
	300	860	0.157	0.843	0.186	-0.074	-0.730	-0.2812	-0.523
	400	800	0.216	0.784	0.275	-0.106	-0.560	-0.2676	-0.398
	500	750	0.265	0.735	0.360	-0.134	-0.443	-0.2757	-0.301
	600	710	0.304	0.696	0.437	-0.157	-0.360	-0.2953	-0.222
	700	680	0.333	0.667	0.499	-0.176	-0.302	-0.3226	-0.155
	800	650	0.363	0.637	0.570	-0.196	-0.244	-0.3432	-0.097
Agar	100	900	0.118	0.882	0.134	-0.054	-0.873	0.07188	-1.000
	200	840	0.176	0.824	0.213	-0.084	-0.670	-0.0555	-0.699
	300	800	0.216	0.784	0.275	-0.106	-0.560	-0.1427	-0.523
	400	750	0.265	0.735	0.360	-0.134	-0.443	-0.1788	-0.398
	500	700	0.314	0.686	0.458	-0.164	-0.339	-0.2020	-0.301
	600	660	0.353	0.647	0.546	-0.189	-0.263	-0.2304	-0.222
	700	620	0.392	0.608	0.645	-0.216	-0.191	-0.2518	-0.155
	800	600	0.412	0.588	0.701	-0.231	-0.154	-0.2882	-0.097
Chitosan	100	810	0.206	0.794	0.259	-0.100	-0.586	0.31387	-1.000
	200	760	0.255	0.745	0.342	-0.128	-0.466	0.10551	-0.699
	300	690	0.323	0.677	0.477	-0.169	-0.321	0.03208	-0.523
	400	645	0.368	0.632	0.582	-0.199	-0.235	-0.0362	-0.398
	500	585	0.426	0.574	0.742	-0.241	-0.129	-0.0696	-0.301
	600	530	0.480	0.520	0.923	-0.284	-0.035	-0.0969	-0.222
	700	480	0.529	0.471	1.123	-0.327	0.050	-0.1216	-0.155
	800	440	0.569	0.431	1.320	-0.365	0.121	-0.1480	-0.097

Arabic gum	100	870	0.147	0.853	0.172	-0.069	-0.764	0.16732	-1.000
	200	800	0.216	0.784	0.275	-0.106	-0.560	0.03342	-0.699
	300	720	0.294	0.706	0.416	-0.151	-0.380	-0.0088	-0.523
	400	680	0.333	0.667	0.499	-0.176	-0.302	-0.0796	-0.398
	500	620	0.392	0.608	0.645	-0.216	-0.191	-0.1057	-0.301
	600	570	0.441	0.559	0.789	-0.252	-0.103	-0.1337	-0.222
	700	530	0.480	0.520	0.923	-0.284	-0.035	-0.1639	-0.155
	800	500	0.510	0.490	1.041	-0.310	0.017	-0.1955	-0.097
Cellulose	100	750	0.265	0.735	0.360	-0.134	-0.443	0.42325	-1.000
acetate	200	700	0.314	0.686	0.458	-0.164	-0.339	0.1959	-0.699
	300	650	0.363	0.637	0.570	-0.196	-0.244	0.08279	-0.523
	400	600	0.412	0.588	0.701	-0.231	-0.154	0.01284	-0.398
	500	560	0.451	0.549	0.821	-0.260	-0.085	-0.0448	-0.301
	600	500	0.510	0.490	1.041	-0.310	0.017	-0.0706	-0.222
	700	440	0.569	0.431	1.320	-0.365	0.121	-0.0899	-0.155
	800	400	0.610	0.390	1.564	-0.409	0.194	-0.1178	-0.097
Cellulose	100	840	0.176	0.824	0.213	-0.084	-0.670	0.24551	-1.000
	200	790	0.225	0.775	0.290	-0.111	-0.537	0.05115	-0.699
	300	700	0.314	0.686	0.458	-0.164	-0.339	0.01981	-0.523
	400	650	0.363	0.637	0.570	-0.196	-0.244	-0.0421	-0.398
	500	600	0.412	0.588	0.701	-0.231	-0.154	-0.0841	-0.301
	600	540	0.471	0.529	0.890	-0.276	-0.050	-0.1051	-0.222
	700	500	0.510	0.490	1.041	-0.310	0.017	-0.1375	-0.155
	800	460	0.550	0.450	1.222	-0.347	0.087	-0.1627	-0.097
Carboxymethyl	100	790	0.225	0.775	0.290	-0.111	-0.537	0.35218	-1.000
cellulose	200	750	0.265	0.735	0.360	-0.134	-0.443	0.12222	-0.699
sodium salt	300	680	0.333	0.667	0.499	-0.176	-0.302	0.04532	-0.523
	400	640	0.372	0.628	0.592	-0.202	-0.227	-0.0310	-0.398
(CMC)	500	575	0.436	0.564	0.773	-0.249	-0.112	-0.0590	-0.301
	600	520	0.490	0.510	0.961	-0.292	-0.017	-0.0880	-0.222
	700	460	0.550	0.450	1.222	-0.347	0.087	-0.1050	-0.155
	800	410	0.598	0.402	1.487	-0.396	0.172	-0.1260	-0.097







Figure 6. Temkin adsorption isotherms at 298 K for Cellulose acetate



Figure7. Kinetic adsorption isotherms at 298 K for Cellulose acetate

Table3. Parameters Flory-Huggins, Temkin and kinetic isotherms for polymers at $8M H_3PO_4$, 3cm height and 298 K in case of using undivided cell.

Organic	Models Parameters								
Compounds	Temkin		Flory-Huggins		Kinetic adsorption Isotherm				
	a K		Х	К	у	1/y	K		
Staech	-1.272	16.65	1.235	1.001	0.896	1.116	1.137		
Gelatin	-1.479	12.52	0.862	0.781	1.008	0.992	0.699		
Agar	-1.484	18.00	1.742	0.716	0.816	1.225	0.996		
Chitosan	-1.222	24.48	1.442	1.462	0.797	1.255	1.724		
Cellulose acetate	-1.319	37.83	1.639	1.813	0.700	1.429	2.162		
Cellulose	-1.171	20.38	1.303	1.374	0.863	1.159	1.563		
CMC	-1.205	25.68	1.333	1.608	0.791	1.264	1.840		

Table4. The values of standard free energy of adsorption for polymers using different adsorption isotherms models at $8M H_3PO_4$, 3cm height and 298 K in case of using undivided cell.

Natural polymers additives	- ΔG_{ads} (K.J. mol ⁻¹)						
	Undivided cell						
	Temkin	Flory- Huggins	Kinetic				
Starch	16.92	9.95	10.27				
Gelatin	16.21	9.34	9.06				
Agar	17.11	9.12	9.94				
Chitosan	17.87	10.89	11.30				
Arabic gum	17.22	10.36	10.74				
Cellulose acetate	18.95	11.42	11.86				
Cellulose	17.42	10.74	11.06				
СМС	17.99	11.13	11.46				

Electropolishing of carbon steel in presence of polymers additives under forced convection mass transfere using rotating cylinder electrode

Table (5) gives the limiting current values at different concentration of polymers additives and different rotation speed at 298 K. We found that the limiting current increases by increasing rotation speed, which indicates that, the electropolishing reaction is a diffusion controlled reaction and decrease with increasing polymer molecules **[35]**.

				Starch			Gelatin				
rpm	100	200	300	500	700	rpm	100	200	300	500	700
С				$\mathbf{I}_{\ell}(\mathbf{A})$		С				$\mathbf{I}_{\ell}(\mathbf{A})$	
(ppm)						(ppm)					
0.00	1.19	1.34	1.48	1.75	1.95	0.00	1.19	1.34	1.48	1.75	1.95
100	1.05	1.20	1.36	1.61	1.80	100	1.13	1.27	1.42	1.69	1.89
200	0.98	1.14	1.30	1.56	1.75	200	1.08	1.23	1.37	1.63	1.83
300	0.93	1.10	1.25	1.50	1.71	300	1.04	1.17	1.30	1.57	1.75
400	0.89	1.05	1.21	1.45	1.65	400	1.00	1.14	1.26	1.52	1.71
500	0.85	1.00	1.16	1.39	1.58	500	0.96	1.11	1.24	1.49	1.67
600	0.82	0.96	1.12	1.35	1.54	600	0.93	1.07	1.20	1.45	1.64
700	0.78	0.92	1.06	1.30	1.50	700	0.90	1.05	1.16	1.40	1.60
800	0.75	0.86	1.00	1.25	1.46	800	0.85	1.02	1.12	1.36	1.55
Agar											
		I.	I.	Agar					(Chitosan	
rpm	100	200	300	Agar 500	700	rpm	100	200	300	Chitosan 500	700
rpm C	100	200	300	Agar 500 I _ℓ (A)	700	rpm C	100	200	300	Chitosan 500 I _l (A)	700
rpm C (ppm)	100	200	300	Agar 500 I _t (A)	700	rpm C (ppm)	100	200	300	Chitosan 500 I _l (A)	700
rpm C (ppm) 0.00	100 1.19	200	300 1.48	Agar 500 I₁(A) 1.75	700	rpm C (ppm) 0.00	100 1.19	200	300 1.48	Chitosan 500 Ι _ℓ (A) 1.75	700 1.95
rpm C (ppm) 0.00 100	100 1.19 1.08	200 1.34 1.25	300 1.48 1.40	Agar 500 Iℓ (A) 1.75 1.66	700 1.95 1.87	rpm C (ppm) 0.00 100	100 1.19 0.91	200 1.34 1.06	300 1.48 1.20	Chitosan 500 I _ℓ (A) 1.75 1.45	700 1.95 1.66
rpm C (ppm) 0.00 100 200	100 1.19 1.08 1.01	200 1.34 1.25 1.20	300 1.48 1.40 1.35	Agar 500 Iℓ (A) 1.75 1.66 1.61	700 1.95 1.87 1.82	rpm C (ppm) 0.00 100 200	100 1.19 0.91 0.85	200 1.34 1.06 1.00	300 1.48 1.20 1.15	Chitosan 500 I I.(A) I.75 1.45 I.40	700 1.95 1.66 1.57
rpm C (ppm) 0.00 100 200 300	100 1.19 1.08 1.01 0.96	200 1.34 1.25 1.20 1.14	300 1.48 1.40 1.35 1.28	Agar 500 I_{ℓ} (A) 1.75 1.66 1.61 1.55	700 1.95 1.87 1.82 1.73	rpm C (ppm) 0.00 100 200 300	100 1.19 0.91 0.85 0.80	200 1.34 1.06 1.00 0.96	300 1.48 1.20 1.15 1.10	Chitosan 500 I_{ℓ} (A) 1.75 1.45 1.40 1.35	700 1.95 1.66 1.57 1.50
rpm C (ppm) 0.00 100 200 300 400	100 1.19 1.08 1.01 0.96 0.92	200 1.34 1.25 1.20 1.14 1.10	300 1.48 1.40 1.35 1.28 1.24	Agar 500 I_{ℓ} (A) 1.75 1.66 1.61 1.55 1.50	700 1.95 1.87 1.82 1.73 1.71	rpm C (ppm) 0.00 100 200 300 400	100 1.19 0.91 0.85 0.80 0.77	200 1.34 1.06 1.00 0.96 0.90	300 1.48 1.20 1.15 1.10 1.03	Chitosan 500 I_{ℓ} (A) 1.75 1.45 1.40 1.35 1.30 I_{ℓ}	700 1.95 1.66 1.57 1.50 1.45
rpm C (ppm) 0.00 100 200 300 400 500	100 1.19 1.08 1.01 0.96 0.92 0.89	200 1.34 1.25 1.20 1.14 1.10 1.04	300 1.48 1.40 1.35 1.28 1.24 1.20	Agar 500 I_{ℓ} (A) 1.75 1.66 1.61 1.55 1.50 1.44	700 1.95 1.87 1.82 1.73 1.71 1.64	rpm C (ppm) 0.00 100 200 300 400 500	100 1.19 0.91 0.85 0.80 0.77 0.74	200 1.34 1.06 1.00 0.96 0.90 0.83	300 1.48 1.20 1.15 1.10 1.03 0.97	Chitosan 500 I_{ℓ} (A) 1.75 1.45 1.40 1.35 1.30 1.25	700 1.95 1.66 1.57 1.50 1.45 1.39
rpm C (ppm) 0.00 100 200 300 400 500 600	100 1.19 1.08 1.01 0.96 0.92 0.89 0.85	200 1.34 1.25 1.20 1.14 1.10 1.04 1.00	300 1.48 1.40 1.35 1.28 1.24 1.20 1.15	Agar 500 I_{ℓ} (A) 1.75 1.66 1.61 1.55 1.50 1.44 1.40	700 1.95 1.87 1.82 1.73 1.71 1.64 1.60	rpm C (ppm) 0.00 100 200 300 400 500 600	100 1.19 0.91 0.85 0.80 0.77 0.74 0.70	200 1.34 1.06 1.00 0.96 0.90 0.83 0.75	300 1.48 1.20 1.15 1.10 1.03 0.97 0.92	Chitosan 500 I_{ℓ} (A) 1.75 1.45 1.45 1.30 1.25 1.20	700 1.95 1.66 1.57 1.50 1.45 1.39 1.34
rpm C (ppm) 0.00 100 200 300 400 500 600 700	100 1.19 1.08 1.01 0.96 0.92 0.89 0.85 0.81	200 1.34 1.25 1.20 1.14 1.10 1.04 1.00 0.95	300 1.48 1.40 1.35 1.28 1.24 1.20 1.15 1.10	Agar 500 I_{ℓ} (A) 1.75 1.66 1.61 1.55 1.50 1.44 1.32	700 1.95 1.87 1.82 1.73 1.71 1.64 1.55	rpm C (ppm) 0.00 100 200 300 400 500 600 700	100 1.19 0.91 0.85 0.80 0.77 0.74 0.70 0.66	200 1.34 1.06 1.00 0.96 0.90 0.83 0.75 0.70	300 1.48 1.20 1.15 1.10 1.03 0.97 0.92 0.86	Chitosan 500 I_{ℓ} (A) 1.75 1.45 1.40 1.35 1.25 1.20 1.16	700 1.95 1.66 1.57 1.50 1.45 1.39 1.34 1.30

Table5. Values of limiting current (I_{ℓ}) for different concentrations of polymers at 8M H_3PO_4 , 3cm height, 298 K and different rotation speeds.

Study the behaviour of polymers molecules on carbon steel surface as a corrosion inhibitors using Scanning electron microscope (SEM) examination:

As shown in Figure (8 a-m), the surface morphology before and after electropolishing (EP) of carbon steel in the absence and in the presence of different concentrations of polymers molecules.

The SEM analysis (Fig. a) shows the morphology of steel before EP. The surface is rough and uneven surface and large number of pits with large size and high depth distributed over the surface are seen. (Fig. b) after EP with no additive Only a slight difference in the surface morphology was observed in which cavities and small pits are represented clearly.(Fig. c), (Fig. d) and (Fig. e) show only small change in the morphology of surface with large amount of small pits and deep cavities.

(Fig .g), (Fig .i) and (Fig .f) show very slightly roughness and more homogenous surface Deep cavities are eliminated by filling up, also, grain boundaries are decreased gradually. This behavior may be due to involvement of Arabic gum, Cellulose and Chitosan molecules in the cavities of steel surface so appear more uniform than blank.

(Fig .k) and (Fig .h) the metallic surface seems to be almost not affected by corrosion and some precipitates observed.

In higher concentration of natural polymers (Fig .L) electropolished surface appear uniform, smooth and bright more than (Fig .c) this seems to be due to adsorption of Starch increases.

(Fig.m) show the SEM images for steel surfaces electropolished at the higher temperatures (318 K) it shows uniform, smooth and bright surface to some extent more than at lower temperature (298 K).

On other hand in presence of natural polymers there is an improvement in the surface morphology due to decreased in the corroded area caused by the inhibitor layer covering the electrode surface.



Fig8a. Raw material



Fig8c. After EP 400 ppm Starch



Fig8e. After EP 400 ppm Agar



Fig8g. After EP 400 ppm Arabic gum



 $Fig8b.\, \textit{After EP} (\textit{blank})$



Fig8d. After EP 400 ppm Gelatin



Fig8f. After EP 400 ppm Chitosan



Fig8h. After EP 400 ppm Cellulose acetate



Fig8i. After EP 400 ppm Cellulose







Fig8k. After EP 400 ppm CMC



Fig8m. *After EP at high temperature (318 K)*

Application of polymers additives on corrosion control of carbon steel pipes

A pilot for distillation tower were used for this application .the overhead of this tower used to release hydrocarbons gases which are corrode the material of this overhead (carbon steel).So many different corrosion inhibitors were injected in this overhead and the efficiency of these corrosion inhibitors were evaluated by measured the Concentration of Fe^{+2} (ppm) in the sample that taken from a drain system.

Concentration of Fe^{+2} (ppm) in case of presence and absence of polymers molecules were measured using Atomic absorption spectroscopy .we found that as concentration of polymers additives increases, the positively ferrous ions Fe^{+2} (ppm) will decrease as shown in Table (6) and Figure(9). So this effect will make the ability to control the corrosion in carbon steel and give an indication for these polymers as agood inhibitors.

	1				2							
C	$Fe^{+2} x 10^{-2} (ppm)$											
(ppm)	Starch	Gelatin	Agar	Chitosan	Arabic	Cellulose	Cellulose	СМС				
			_		gum	acetate						
0	987.4	987.4	987.4	987.4	987.4	987.4	987.4	987.4				
100	630.6	870.0	788.9	279.3	495.03	88.60	440.7	151.1				
200	606.9	822.7	760.1	230.06	474.7	79.58	420.4	132.2				
300	570.32	781.0	740.8	198.97	451.5	70.06	370.3	120.5				
400	530.87	740.7	719.6	166.13	420.8	63.99	340.96	109.8				
500	496.23	709.0	697.8	134.09	398.06	55.48	299.1	100.2				
600	450.98	688.9	670.3	110.76	360.87	46.01	270.7	90.05				
700	410.03	634.6	605.1	90.94	290.07	35.72	219.08	70.30				
800	370.55	591.1	565.9	75.81	233.19	20.41	170.11	50.77				

Table6. Values of Fe^{+2} (ppm) for different concentrations of polymers additives as corrosion inhibitors at 298 K.



Figure9. Relation between ferrous ion concentration $Fe^{+2}(ppm)$ and different concentrations of polymers additives as corrosion inhibitors at 298 K.

REFERENCES

- [1] P.A. Jaquet, Trans. Electrochem. Soc., 69, 629 (1936).
- [2] S. Shivareddy, S. E. Bae and S. R. Brankovic, Electrochem. Solid-State Lett. 11, 13 (2008).
- [3] L. D. Burke and R. Sharna, J. Electrochem. Soc. 155, 83 (2008).
- [4] S. P. Wu, L. Jiao, J. Ni, Z. O. Zeng and S. Liu, Intermetallics 15, 1316 (2007).
- [5] E. M. Sherif and S.-M. Park, Corros. Sci. 48, 4065 (2006).
- [6] A.M. Awad, N.A. Abdel Ghany and T.M. Dahy, J. Appl. Surf. Sci., 256, 4370 (2010).
- [7] T.T. Qin, J. Li, H.Q. Luo, M. Li and N.B. Li, Corros. Sci., 53, 1072 (2011).
- [8] Revised Draft SGEIS on the Oil, Gas and Solution Mining Regulatory Program. Dep. of Environment Protection, NY, 5.4.3.1. Chemical Categories and Health Information, (2011).
- [9] U. Ateya, B.E. EL-Anadouli and F.M. EL-Nizamy, Corros. Sci., 24,509,(1984).
- [10] S. Abd El-Wanees and E.E. Abd El-Aal, Corros. Sci., 52, 338 (2010).
- [11] F. Jia-Jun, S-n. Li, Y. Wang, L-H. Cao and L. Lu-de, J. Mater. Sci. 46, 6255 (2010).
- [12] J. C. Da Rocha, D. C. P. Gomes and E. D'Elia, Corros. Sci. 52, 234 (2010).
- [13] V. R. Saliyan and A. V. Adhikari, Bull. Mater. Sci. 31, 699 (2008).
- [14] A. A. Taha, A. M. Ahmed, H. H. Abdel Rahman and F.M. Abouzeid, Appl. Surf. Sci. 277, 155 (2013).
- [15] A. S. Yaro, A. A. Khadom and H. F. Ibraheem, Anti-Corros. Methods Mater. 58, 116 (2011).
- [16] T. Juuti, S. Uusikallio, A. J. Kaijalainen, E. Heinonen, N. T. Tun, D. A. Porter, "The Effect of Sample Preparation on The Microstructure of Austenitic-Ferritic Stainless Steel", Materials Science Forum, Vol.879, PP.873-878, 2017.
- [17] Alykov N. M., Tyrkov A. G., Pichugina E. A., Tyrkova E. A.: Inhibitors of acidic corrosion for protection of oil and gas pipelines. RU patent 2365679, (2009).
- [18] MacDonald D. D., Roberts B., Hyne J. B.: The corrosion of carbon steel by wet elemental sulphur. Corrosion Science 1987, 18, pp. 411–425.
- [19] F. Mounir, S. El Issami, Lh. Bazzi, R. Salghi, L. Bammou, A. Chihab Eddine and O. Jbara, Int. J. Res. Rev. Appl. Sci. (IJRRAS). 13, 574 (2012).
- [20] S. Arrhenius, Z. Phyzik Chem., 285 (1887).
- [21] M. A. Shaker and H.H. Abdel Rahman, Am. J. Appl. Sci. 4, 554 (2007).
- [22] A. Ketsetzi, A. Stathoulopoulou and K. D. Demadis, Desalination 223,487 (2008).
- [23] C. O. Akalezi, C. K. Enenebaku and E. E. Oguzie, Int. J. Ind. Chem. 3, 1 (2012).
- [24] H.M.A. Soliman and H.H. Abdel Rahman, J. Brazil. Chem. Soc., 17, 705 (2006).
- [25] G. M. El-Subruiti, A. M. Ahmed and M. G. Koretam, Bull. Electrochem. 19, 185 (2003).

International Journal of Advanced Research in Chemical Science (IJARCS)

- [26] M. A. Shaker and H.H. Abdel Rahman, Am. J. Appl. Sci. 4, 554 (2007).
- [27] G. Moretti, F. Guidi, Corros. Sci., 44, 1995, (2002).
- [28] M. Shah, A. A. Rahim, S. A. Hamid and S. Yahya, Int. J. Electrochem. Sci. 8, 2140 (2013).
- [29] E. E. Oguize, B. N. Okolue , C. L Ogukwl Bull Electrochem. 20, 421, (2004).
- [30] E. A. Ayobe, S. F. Abaza, S. M. Seleim and A. M. Ahmed, Indian J. Chem. Technol. 19, 322 (2012).
- [31] D. A. Awizar, N. K. Othman, A. Jala, A. R. Daud, I. Abdul Rahman and N. H. Al-hardan, Int. J. Electrochem. Sci. 8, 1759 (2013).
- [32] B. M. Mistry, N. S. Patel, S. Sahoo and S. Jauhari, Bull. Mater. Sci. 35, 459 (2012).
- [33] H.H. Abdel Rahman, A.H.E. Moustafa and M.K. Awad, Int. J. Electrochem. Sci. 7, 1266 (2012).
- [34] A. M. Ahmed, S. M. Abd El-Haleem, M. M. Darweesh and A. E. El-Naggar, Bull. Electrochem. 19, 489 (2003).
- [35] N.M. El-Mallah, A.M. Ahmed and L.F. Gado, J. Dispers. Sci. Technol., 31, 1579 (2010).

Citation: H. F.M El-shamy, "Study the Mettalic Finishing and Corrosion Control Processes for Carbon Steel over Head Pipes Using Polymer Molecules International Journal of Advanced Research in Chemical Science (IJARCS)", International Journal of Advanced Research in Chemical Science (IJARCS), vol. 4, no. 8, pp. 31-44, 2017. http://dx.doi.org/10.20431/2349-0403.0408005

Copyright: © 2017 Authors. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

International Journal of Advanced Research in Chemical Science (IJARCS)