

Effects Chemical Treatments and Stratification on Seedlings Emergence of Persian Parrotia (*Parrotia Persica* (DC.) and Assessment of Genetic Diversity in its Seedlings

Hamid Reza Karimi*¹, Elham Sadeghi-Seresht², Shirin Nasrolahpour-Moghadam²,
Sepideh Soleimani², Homayoun Farahmand³, Mohamad Sadegh Jome-Yazdian²

¹Associate Professor of Department of Horticultural Science, Faculty of Agriculture, Vale-e- Asr University of Rafsanjan, Iran.

²MSc Student of Department of Horticultural Science, Faculty of Agriculture, Vale-e- Asr University of Rafsanjan, Iran.

³Department of Horticultural Science, Faculty of Agriculture, Shahid Bahonar University of Kerman, Kerman, Iran.

***Corresponding Author:** Hamid Reza Karimi, Associate Professor of Department of Horticultural Science, Faculty of Agriculture, Vale-e- Asr University of Rafsanjan, Iran.

Abstract: Persian parrotia (*Parrotia persica* C.A. Mey.) is native to northern Iran and Azerbaijan, along the Caspian Sea. Persian parrotia is an important tree species in Hyrcanian forests and also well known for its landscape values and the wood is also used by wood turners and for weaving shuttles, telephone poles. Very limited agronomic information exists regarding the cultivation of Iron tree as a industrial crop. In this study we investigated effects of cold stratification (25, 50 and 75 days) in combination with putrescine (5, 10 and 15 mM), BA (1, 2 and 3 mM), KNO₃ (25, 50 and 75 mM) and warm stratification (42 days) on seed germination and emergence of *P. persica* and assessment of genetic diversity in it seedlings using RAPD markers. The results of germination treatments showed that the highest emergence percentage was obtained with 42 days warm moist chilling followed by 50 days cold stratification. To assessment of genetic diversity, 18 RAPD primers were used. The results indicated that 6 out of 18 primers produced no band on the studied genotypes and the rest primers (12) produced 92 bands overall. The average polymorphism bands were 7.66%. The lowest number of bands was three bands with 66.6% polymorphism which obtained with OPB-10 primer and the highest number of polymorph bands was 12 bands with 100% polymorphism gained with OPD-05 primer. The highest resolving power (0.48 and 0.47) was found with OPA-10 and OPB-10 primers, respectively. The lowest resolving power (0.23) was related to OPE-06 and TIBMBC-13 primers. The molecular results showed that OPD-05 and OPA-10 and OPA09 primers can use for ongoing genetic variation studies in Persian parrotia.

Keywords: *Parrotia persica*, seed germination, treatment.

1. INTRODUCTION

The family Hamamelidaceae comprises 30 genera and 144 species and has been the subject of a great attention due to its phylogenetic position and extensive fossil records (Zhou et al. 2001; Andrew 1997; Zhi-yun and An-ming 1995). Persian parrotia, Persian ironwood, Iron tree or Ironwood tree (*Parrotia persica* C.A. Mey.) was described by C.A. Meyer in 1831 and named in honor of F.W. Parrot, a German physician, naturalist and explorer (More and White 2003; Li and Tredici 2006; Andrews 2007). Persian parrotia is native to northern Iran and Azerbaijan, along the Caspian Sea, a phytogeographic region consists of northern Iran (Gilan, Mazandarn, Gorhgan, north-west Khorasan) and Talish in Azerbaijan (Sabeti 1994; Mozaffarian 2003; Mozaffarian 2005; Ahanjan 2007; Andrews 2007). The species with some its cultivars are now found in many famous botanic gardens around the world (Andrews 2007). Persian ironwood is a highly ornamental tree or large shrub that can grow to 20-25 m and has been in cultivation since 1840 (More and White 2003; Andrew 2007). It makes a round-year feature with red or

brown bark patches in winter, lustrous green leaves in summer following by quite outstanding color change in autumn which makes it ideal as specimen, accent or street tree (More and White 2003; Gilman 2014). It is also a tough species that tolerates drought, heat, wind and cold (Gilman 2014). The species is also one of the most lime tolerate ones in Hamamelidaceae (Andrew 2007).

This species has some local Persian names including "Anjeeli" and is praised for many characteristics including its fabulous autumn color change particularly in mixed woods and used in landscape and botanical gardens in Iran, as well. The wood is extremely hard and durable and this is why it is called iron tree. The charcoal made from it is highly valued. The wood is also used by wood turners and for weaving shuttles, telephone poles or other uses in Iran and Azerbaijan (Sabeti 1994; Andrew 1997). Many herbalists use Persian ironwood in the treatment of various fevers and respiratory infections in Iran due to its flavonoids compounds and is also used for food coloring and food flavoring (Ahanjan et al. 2007). There are various kinds of dormancy in which physiological dormancy may be decreased by cold or warm stratification or application of hormones (Takos 2001; Zhou et al. 2003; Garcia- Gusano et al. 2004; Rehman et al. 2000; Duan et al. 2004; Fang et al. 2006; El- Refaey 2014). Plant growth regulators such as GA₃ (Gibberellic acid) and BA (Benzyl adenine) and chemical compounds including KNO₃ (Potassium nitrate) and putrescine may improve seed germination and/or early seedling growth in plant species (Bryan and Seiler 1991; Matilla 1996; Strik et al. 2005; El-Tohamy et al. 2008; Tzortzakis 2009). A combination treatment of alternating temperatures (25/15 or 23/11°C) for 6 months followed by 5 °C for 3 months was successful in overcoming seed dormancy in *Taxus mairei* (Chien et al. 1998). Warm stratification followed by cold stratification has also been applied in some *Rosa* taxa (Alp et al. 2009) and *Styrax japonicas* (Alp et al. 2009) to overcome seed dormancy. Warm stratification has also been reported as a requirement for breaking dormancy in seeds with intermediate physiological dormancy such as *Empetrum hermaphroditum* (Baskin et al. 2002). Seed germination of Japanese Stewartia (*Camelia japonica*) was promoted by GA₃ treatment and warm and cold stratification (Olesksak and Struve 1999). The application of GA₃ during and after stratification significantly increased the length, trunk diameter, internodes length, leaf area and fresh and dry weight of seedlings of two species of *Pistacia* (Rahemi and Baninasab 2000). According to Airi et al. (2009), treating the seeds of *Hippophae salicifolia* with GA₃ shortened mean germination time and increased germination percentage. GA₃ treatment followed by warm stratification for 3 months and 7 months of cold stratification has been reported as a practical procedure for seed germination in *Stewartia pseudocamellia* (Brian et al. 1999). In *Prunus avium*, treating the seeds with 7500 ppm KNO₃ after 120 days of stratification was very effective and resulted in 65% germination of seeds with coat (Cetnibas and Koyunncu 2006). The positive effect of potassium nitrate has reported for the genus *Rubus* (Wada and Reed 2011). Polyamines are implicated in seed germination and their concentrations altered by the process of stratification (Matilla 1996). Application of polyamins (Spd, Spm and Put) on pistachio, increased both seed germination percentage and mean germination rate and seedling growth (Sedaghat and Rahemi 2011). Chapurro et al. (1988) studied effects of BA on peach seeds germination and reported that the highest germination was obtained by 24 hr exposure to 50-100 mg L⁻¹ of BA. Osman Sarihan et al. (2005) investigated the effects of GA₃ and KNO₃ on *Plantago lanceolata* seeds and reported the positive effects of these compounds on seed germination. Some members of Hamamelidaceae require warm and cold stratification treatments to germinate successfully. Seed germination of 17 species of this family was investigated at Batumi Botanical Garden using scarification and stratification treatments and a long period of dormancy reported for some species including *Parrotia persica* (Metreveli and Bregvadze 2007). *P. persica* is propagated by layering, grafting on *Hammamelis virginiana*, root off-shoots or root suckers, cutting under mist and micropropagation (Sabeti 1994; Andrew 1997; Hartman et al. 2011). It can also be raised by seed (Hartman et al. 2011). The natural regeneration of the species through seed is also very scanty. *Parrotia persica* is a slow- growing tree, preservation of the present populations and a genetic database information for future purposes are of vital importance. Consequently, optimizing protocols for genetic assessment and adoption the efficient means to pave the ground for breeding and biotechnological goals are very decisive. On the other hands, there are just few reports (Yosefzadeh et al. 2010, Sattarian et al. 2011) covering the phylogenetic relationship among Persian parrotia populations in Iran merely based on morphological markers.

Recent years have witnessed increasingly rapid development of molecular phylogenetics and systematics. This is due to the development of new diverse methods of analysis of molecular DNA markers (Grechko 2002). To investigate the phylogenetic relationship among the populations of each plant with DNA markers, the first step is the adoption of efficient primers. In the case of *Persia parrotia*, no report is available about using molecular markers for genetic variation assessment. There are a few reports on seed germination of iron wood tree and its genetic variation assessment. Thus, the objective of this study was to investigate effect of cold and warm stratification and some chemical compounds on germination of Persian parrotia and assessment of genetic diversity in its seedlings using RAPD marker as an important forest species and suitable for landscape.

2. MATERIAL AND METHODS

2.1. First Experiment (Seed Germination Treatments)

The mature seeds with dark brown colour were collected from Noor forest in the western

Hyrceanian forest of Iran. The geographical location of the seed collection was 52° E and 36° N with annual mean of temperature and precipitation 16.5° C and 850 mm respectively (Sattarian et al. 2011). The seeds were collected from 20 trees and were stored at 4 ° C to start testing. The treatments were done on the same year. The seeds were surface sterilized with sodium hypochlorite (1%) for 5 min and kept in distilled water for 24hr. For cold stratification treatment, the seeds were mixed with perlite (1:3 seed/perlite) as medium and put in refrigerator at 4±1 C° for 25, 50 and 75 days (Baninasab and Rahemi 2008). At the end of stratification period, the seeds were taken out and treated with KNO₃ (25, 50 and 75 mM), putrescine (5, 10 and 15 mM) (Sedaghat and Rahemi 2001) and BA (1, 2 and 3 mM) (Khalil and Al-Eisawi 1998) and were sown in pots with perlite medium. For warm stratification, seeds were treated at 25±1° C for 42 days in moistened perlite (Bujarska-Borkowska 2002). After warm stratification, the seeds were sown in pots with perlite medium and kept in greenhouse at 24± 2 °C. The present study was done in four parts that the first part was putrescine treatment with cold stratification, the second and third parts were KNO₃ and BA treatments respectively with cold stratification and final part was warm stratification treatment with cold stratification. In all treatments, control was 25 days cold stratification. Each part of study had 10 treatments except to final part that was contained 4 treatments. Each treatment had four pots as replication. In total 100 seeds were used in each treatment. Pots were irrigated during experiment period with Hogland solution. Seedling emergence percentage was recorded 30 days after sowing. All treatments of this experiments are presented in Tables 2, 3, 4 and 6.

Seedling emergence percentage and emergence rate were calculated 30 days after seed sowing using the following equations (Salim Azad et al. 2011; Bian et al. 2013). Meanwhile, some growth parameters such as leaf area, leaf number, stem length and shoot and root dry mass were measured.

Emergence rate = $\frac{\text{number of seedlings/days to first count} + \dots + \text{number of seedlings/days to final count}}{\dots}$

Emergence (%) = $a \times 100/N$

Where, E is emergence percentage; a is the number of produced seedlings after 30 days, and N is the total number of planted seeds. Leaf area of three youngest, fully expanded leaves from the terminal shoots were measured with a portable leaf area meter (LI-COR 3000, Lincoln, Neb.) in 90 days after seed sowing. At the end of the experiment; 90 days after seed sowing; plants were cut at the pot level and roots were washed and separately oven-dried at 70 °C for measurement of root and shoot dry weight. In part of experiment design was as completely randomized design (CRD) with four replications each contained 25 seeds per pot (100 seeds per each treatment). Analysis of variance (ANOVA) was performed using the SAS version 9.4 software's for each part of experiment. If ANOVA determined that the effects of the treatments were significant ($P \leq 0.05$), then the means were compared by Tukey's multiple range test.

2.2. Second Experiment (Molecular Evaluation)

Eighteen genotypes (seedling) produced from the first study were used in this study, but 5 did not produce any bands by used primers or did not amplify clear products. In total, 13 genotypes which produced good and reproducible bands were used for further analysis. The leaf samples were washed three times in sterile distilled water, frozen in liquid nitrogen and kept at -20°C until used. Genomic DNA was extracted using the CTAB method of Doyle and Doyle (1987) with minor modifications (Karimi et al. 200; Karimi and Sadeghi Seresht 2015). One gram of needle was ground in liquid nitrogen and mixed with 6 mL of CTAB buffer (100 mM Tris- HCl, 1.4 M NaCl, 20 mM EDTA, 2% CTAB, 2% polyvinylpyrrolidone, 0.2% β -mercaptoethanol, 0.1% $\text{Na}_2\text{S}_2\text{O}_5$). The samples were then incubated at 65°C for 1 h, following by extraction with an equal volume of chloroform- isoamylalcohol (24/1). The aqueous phase was recovered and mixed with an equal volume of cold isopropanol and kept at -24°C for 24 hour. The precipitated nucleic acids were recovered by centrifugation at 1000 rpm for 2 min, washed with ammonium acetate in 76% ethanol, dried and resuspended with double distil water. DNA concentration was estimated spectrophotometrically and confirmed by electrophoresis in 8% agarose gels using known concentration of bacteriophage lamda DNA (CinnaGen, Tehran, Iran)

Table1. List of used primers to assessment of genetic diversity in iron wood genotypes (seedlings)

Row	Primer	Primer sequence
1	OPD-03	GTCGCCGTC
2	OPD-05	TGAGCGGACA
3	OPD-06	ACCTGAACGG
4	OPD-07	TTGGCACGGG
5	OPD-08	GTGTGCCCCA
6	OPD-20	ACCCGGTCAC
7	OPAD-02	CTGAACGCTG
8	OPA-09	GGGTAACGCC
9	OPA-08	GTGACGTAGG
10	OPA-10	GTGATCGCAG
11	OPB-10	CTGCTGGGAC
12	OPC-02	GTGAGGCGTC
13	OPE-06	AAGACCCCTC
14	OPG-02	GGCACTGAGG
15	OPZ-10	CCGACAAACC
16	OPAE-10	CTGAAGCGCA
17	TIBMBB-12	GTGTGCCCCA
18	TIBMBC-13	TCGGTGAGTC

At the beginning of the experiment, 72 Operon 10-mer primers (Operon Technologies, Alameda, CA, USA) and 100 TIB 10-mer primers (TIBMOLBIOL, Berlin, Germany), were tested on three genotypes and 18 primers was selected for the next step in this study (Table 1). Polymerase chain reactions (25 μ) each contained 10 ng template DNA, 1 \times PCR buffer (CinnaGen, Tehran, Iran), 0.875 mM MgCl_2 , 200 μM each of dNTPs, 0.2 μM each decamer primer, and 1 unit of *Taq* DNA polymerase (CinnaGen, Tehran, Iran). Amplification reactions were performed in termocycler (iCycler, Bio Rad, Hercules, CA, USA) programmed as follow: 94°C for 4 min, followed by 35 cycle of 92°C for 1 min, 37°C for 1 min, 72°C for 2 min and a final extension at 72°C for 5 min. Amplified products were separated by electrophoresis in 1.5% (w/v) agarose gels in Tris-borate-EDTA (TBE) buffer (89 mM Tris, 89 mM Boric acid, 2 mM EDTA. Na_2 , pH=8.0), visualized by ethidium bromide staining and photographed under UV light with a Gel Doc system (UVP: Bio Doc, Upland, CA, USA). The Simple Matching Coefficient similarity matrix was calculated using numerical taxonomy and multivariate analysis system NTSYSpc Ver 2.11 (Rohlf 2004) and the dendrogram produced using the UPGMA. For each primer, Polymorphism Information Content (PIC) was measured (Farahmand et al. 2015).

3. RESULTS

3.1. First Experiment (Seed Germination)

3.1.1. Effects of Cold Stratification and Putrescine on Emergence

Table 2. Effects of cold stratification and putrescine on seedling emergence and growth parameters of Iron tree.

Treatments	Leaf area (cm ²)	No of leaf	Stem length (cm)	Shoot dry mass (gr)	Root dry mass (gr)	Emergence (%)	Emergence rate (seedling per day)
Control (25 days cold stratification)	0.38 c	3.60 a	4.0 a	0.13 h	0.06 d	20 e	1.2 d
Cold stratification (25 days) + KNO ₃ (25 mM)	2.45 b	4.80 a	5.38a	0.21 c	0.15 a	40 b	1.95 b
Cold stratification (25 days) + KNO ₃ (50 mM)	3.54 b	5.0 a	5.50 a	0.24 b	0.11 c	35 c	1.7 c
Cold stratification (25 days) + KNO ₃ (75 mM)	4.30 b	4.60 a	5.80 a	0.17 e	0.12 b	25 d	1.5 cd
Cold stratification (50 days) + KNO ₃ (25 mM)	2.40 b	4.20 a	4.30 a	0.18 d	0.12 b	45 a	2.7 a
Cold stratification (50 days) + KNO ₃ (50 mM)	2.33 b	3.40 a	4.80 a	0.14 g	0.11 c	40 b	1.95 b
Cold stratification (50 days) + KNO ₃ (75 mM)	4.26 b	3.60 a	4.0 a	0.15 f	0.06 d	35 c	1.7 c
Cold stratification (75 days) + KNO ₃ (25 mM)	3.90 b	6.60 a	7.30 a	0.18 d	0.06 d	20 e	0.95 e
Cold stratification (75 days) + KNO ₃ (50 mM)	4.60 b	6.0 a	7.75 a	0.25 a	0.05 e	15f	0.70 f
Cold stratification (75 days) + KNO ₃ (75 mM)	12.91a	5.50 a	6.45 a	0.13h	0.02 f	10 g	0.45 g

In each column, means with the similar letter (s) are not significantly different at 5% level of probability using Tukey's multiple range test.

The application of putrescine up to 10 mM increased emergence percentage and rate when it was used after 50 days stratification, so that, the highest emergence rate was obtained in combined treatment of 50 day stratification and 5 mM putrescine in compared to control. Seedling emergence percentage and rate were decreased when the seeds receiving 75 days stratification, treated with 10 and 15 mM putrescine (Table 2).

3.1.2. Effects of Cold Stratification and Putrescine on Seedling Growth Parameters

All treatments increased leaf area compared to control significantly, but no significant difference was found among the treatments. The number of leaf was unaffected by the interaction of cold stratification and putrescine and there was no difference among the treatments. Stem length was increased in all treatments except 75 days stratification and 15 mM putrescine. Shoot and root dry weight was increased at all concentrations of putrescine after 25 and 50 days of stratifications but these parameters were significantly decreased compared to control treatment, when putrescine (15 mM) was applied after 75 days of stratification. The highest shoot and root dry mass were obtained with 5 mM putrescine after 75 and 25 days of stratification (Table 2).

3.1.3. Effects of Cold Stratification and KNO₃ on Emergence

All concentrations of KNO₃ increased emergence percentage and rate in comparison to control after 25 and 5 days of stratification but emergence percentage and rate were decreased when KNO₃ applied after 75 days of stratification. The highest seedling emergence was obtained with 50 days stratification and 25 mM KNO₃ (Table 3).

3.1.4. Effects of Cold Stratification and KNO₃ on Seedling Growth Parameters

Table3. *Effects of cold stratification and KNO₃ on seedling emergence and growth parameters of Iron tree.*

Treatments	Leaf area (cm ²)	No of leaf	Stem length (cm)	Shoot dry mass (gr)	Root dry mass (gr)	Emergence (%)	Emergence rate (seedling per day)
Control (25 days cold stratification)	0.38 c	3.60 a	4.0 a	0.13 h	0.06 d	20 e	1.2 d
Cold stratification (25 days) + KNO ₃ (25 mM)	2.45 b	4.80 a	5.38a	0.21 c	0.15 a	40 b	1.95 b
Cold stratification (25 days) + KNO ₃ (50 mM)	3.54 b	5.0 a	5.50 a	0.24 b	0.11 c	35 c	1.7 c
Cold stratification (25 days) + KNO ₃ (75 mM)	4.30 b	4.60 a	5.80 a	0.17 e	0.12 b	25 d	1.5 cd
Cold stratification (50 days) + KNO ₃ (25 mM)	2.40 b	4.20 a	4.30 a	0.18 d	0.12 b	45 a	2.7 a
Cold stratification (50 days) + KNO ₃ (50 mM)	2.33 b	3.40 a	4.80 a	0.14 g	0.11 c	40 b	1.95 b
Cold stratification (50 days) + KNO ₃ (75 mM)	4.26 b	3.60 a	4.0 a	0.15 f	0.06 d	35 c	1.7 c
Cold stratification (75 days) + KNO ₃ (25 mM)	3.90 b	6.60 a	7.30 a	0.18 d	0.06 d	20 e	0.95 e
Cold stratification (75 days) + KNO ₃ (50 mM)	4.60 b	6.0 a	7.75 a	0.25 a	0.05 e	15f	0.70 f
Cold stratification (75 days) + KNO ₃ (75 mM)	12.91a	5.50 a	6.45 a	0.13h	0.02 f	10 g	0.45 g

Interaction of cold stratification and KNO₃ had no significant effects on leaf number and stem length and there was no significant difference between treatments and control. Leaf area was significantly increased in all treatments in comparison to control. The highest leaf area was gained with 75 days stratification and 75 mM KNO₃. In all cold stratification treatments with the exception of 75 days stratification and 75 mM KNO₃, shoot dry mass was increased compared to control. Root dry mass was increased when KNO₃ was used after 25 and 50 days of stratification but it was reduced when KNO₃ (50 and 75 mM) applied after 75 days stratification (Table 3).

3.1.5. Effects of Cold Stratification and BA on Emergence

Emergence percentage and rate were increased compared to control when BA was used after 50 days of stratification treatment. The highest emergence was observed with 2 mM BA. Seedling emergence was decreased when BA was applied after 75 days of stratification (Table 4).

3.1.6. Effects of Cold Stratification and BA on Seedling Growth Parameters

Table4. *Effects of cold stratification and BA on seedling emergence and growth parameters of Iron tree.*

Treatments	Leaf area (cm ²)	No of leaf	Stem length (cm)	Shoot dry mass (gr)	Root dry mas (gr)	Emergence (%)	Emergence rate (seedling per day)
Control (25 days cold stratification)	0.38f	3.60 a	4.0 a	0.13de	0.06 de	20 e	1.2 d
Cold stratification (25 days) + BA (1mM)	1.25e	4.0 a	5.0 a	0.11 e	0.09 d	22 d	1.1 d
Cold stratification (25 days) + BA (2 mM)	1.25 e	4.90 a	5.60 a	0.22 b	0.11 c	19 e	0.96 e
Cold stratification (25 days) + BA (3 mM)	2.68 d	3.50 a	4.23 a	0.33 a	0.23 a	9 g	0.45 j
Cold stratification (50 days) + BA (1mM)	4.37 b	2.25 a	3.60 a	0.09 f	0.02 e	29 b	1.45 b
Cold stratification (50 days) + BA (2 mM)	3.56 c	5.0 a	3.40 a	0.15 d	0.02 e	34 a	1.7 a

Effects Chemical Treatments and Stratification on Seedlings Emergence of Persian Parrotia (*Parrotia Persica* (DC.) and Assessment of Genetic Diversity in its Seedlings

Cold stratification (50 days) + BA (3 mM)	2.28 d	4.0 a	3.54 a	0.18 c	0.12 b	24 c	1.5 c
Cold stratification (75 days) + BA (1mM)	4.0 bc	6.0 a	6.60 a	0.11 e	0.01f	14 f	0.7 f
Cold stratification (75 days) + BA (2 mM)	4.05 bc	6.0 a	6.60 a	0.11 e	0.01f	14 f	0.45 g
Cold stratification (75 days) + BA (3 mM)	5.70 a	8.0 a	9.0 a	0.25 c	0.02 e	9 g	0.45 g

In each column, means with the similar letter (s) are not significantly different at 5% level of probability using Tukey's multiple range test.

Leaf area was increased in all treatments by BA. Leaf number and stem length were not affected by BA treatment, and there was no significant difference compared to control.

The highest leaf area was observed in 75 days stratification and 3 mM BA. Shoot dry mass was increased at 3 mM BA in all stratification levels. Root dry weight was increased at 3 mM BA after 25 and 50 days stratification, but was not affected after 75 days stratification (Table 4).

3.1.7. Effects of Putrescine, KNO₃ and BA on Emergence

Mean comparison of the solely effects of chemical treatments on emergence percentage and rate indicated that seedling emergence and rate were affected by kind of chemical material, so that the highest emergence percentage and rate were observed with KNO₃ treatment. There were not significantly difference between BA and putrescine related to seedling emergence (Table 5).

3.1.8. Effects of Putrescine, KNO₃ And BA On Seedling Growth Parameters

Table5. Comparison of KNO₃, BA and putrescine tratments seedling emergence and growth parameters of Iron tree.

Treatments	Leaf area (cm ²)	No of leaf	Stem length (cm)	Shoot dry mass (gr)	Root dry mass (gr)	Emergence (%)	Emergence rate (seedling per day)
Putrescine	3.26 b	4.65 b	5.03 b	0.19 a	0.14 a	22.70 b	1.30 b
KNO ₃	2.73 b	3. 59 c	3.90 c	0.13 b	0.07 b	34 a	1.70 a
BA	4.82.a	6.57 a	7.43 a	0.03 b	0.03 c	13 b	0.67 c

In each column, means with the similar letter(s) are not significantly different at 5% level of probability using Tukey's multiple range test.

Effects of chemical treatments on measured parameters showed that there was no significant difference between chemical treatments for leaf number, stem length and root dry mass whereas the highest shoot dry mass was obtained with putrescine (Table 5).

3.1.9. Effects of Cold and Warm Stratification on Emergence

Table6. Effects of warm and cold stratification on seedling emergence and growth parameters of Iron tree.

Treatments	Leaf area (cm ²)	No of leaf	Stem length (cm)	Shoot dry mass (gr)	Root dry mass (gr)	Emergence (%)	Emergence rate (seedling per day)
Control (25 days cold stratification)	0.38 d	3.60 b	4.0 c	0.13 c	0.06 a	20 d	1.2 c
Warm stratification (42 days) + without Cold stratification	6.01 c	6.0 a	8.32 b	0.11 d	0.03 b	29 c	1.45 d
Warm stratification (42 days) + cold stratification (25 days)	11.01 b	6.60 a	10.0 ab	0.31 b	0.05 b	34 b	1.7 b
Warm stratification (42 days) + cold stratification (50 days)	16.34 a	7.60 a	11.90 a	0.34 a	0.06 a	44 a	2.2 a

In each column, means with the similar letter(s) are not significantly different at 5% level of probability using Tukey's multiple range tests.

Warm stratification increased seedling emergence in comparison to control. A synergistic effect was found combining warm and cold stratification treatments. The highest emergence percentage in the entire experiment was observed when seeds received 50 days of cold stratification subjected to 42 days of warm stratification (Table 6).

3.2. Second Experiment (Molecular Evaluation)

3.2.1. Discriminative Ability and Efficiency of Primers in Polymorphism

Table7. The number of bands, percentage of polymorphism bands produced by used primers in iron tree genotypes.

Row	Primer	Number of produced bands	Percentage of polymorphism *	RP	PIC
1	OPA-08	7	71.42	1.31	0.44
2	OPB-10	3	66.6	0.87	0.47
3	OPD-03	4	75.0	0.76	0.41
4	OPD-05	12	100	0.93	0.41
5	OPE-06	5	80.0	0.27	0.23
6	OPG-02	10	100	0.49	0.33
7	OPZ-10	8	100	0.86	0.45
8	OPA-09	10	100	0.83	0.48
9	OPAE-10	9	88.8	0.42	0.26
10	OPA-10	11	100	0.37	0.25
11	TIBMBB-12	8	87.5	0.48	0.33
12	TIBMBC-13	5	100	0.27	0.23
Total		92	-	7.33	
Mean		7.66	89.11	0.61	

*(b/a)×100

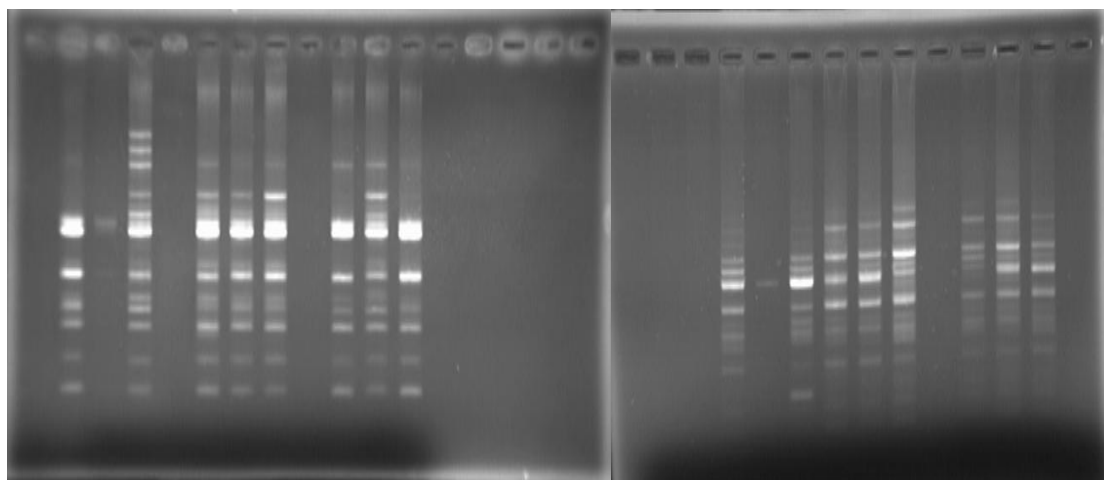


Figure1. RAPD amplified with the arbitrary primer OPD-05 using DNAs of 18 iron tree genotypes.

To investigate the amount of polymorphism among the genotypes of Persian parrotia, only the bands produced by 12 primers were included and the data of 6 primers were not used for analysis (Table 7). In general, 92 bands with average of 7.66 bands per primer were produced by the applied primers. Most of the primers (12 out of 18) exhibited polymorphism. The number of multiplied bands ranged from 3-12 among the 12 primers. The highest band number was produced by OPD-05 (12) and OPA-10 (11) primers (Fig 1). The overall resolving power of primers was 7.33 and 0.61 for each primer. The highest resolving power (0.93) among the used primers was related to OPD-05 primer. Based on the results, the highest polymorphic information content (PIC) 0.48, 0.47 and 0.45, were identified with OPA-09, OPB-10 and OPZ-10 primers, respectively. Thus, the polymorph bands produced in response to PCR were higher in these primers compared to others and these four primers indicated the genetic distance among Persian parrotia more efficiently.

3.2.2. The Amount of Similarity and Cluster Analysis Between the Studied Genotypes

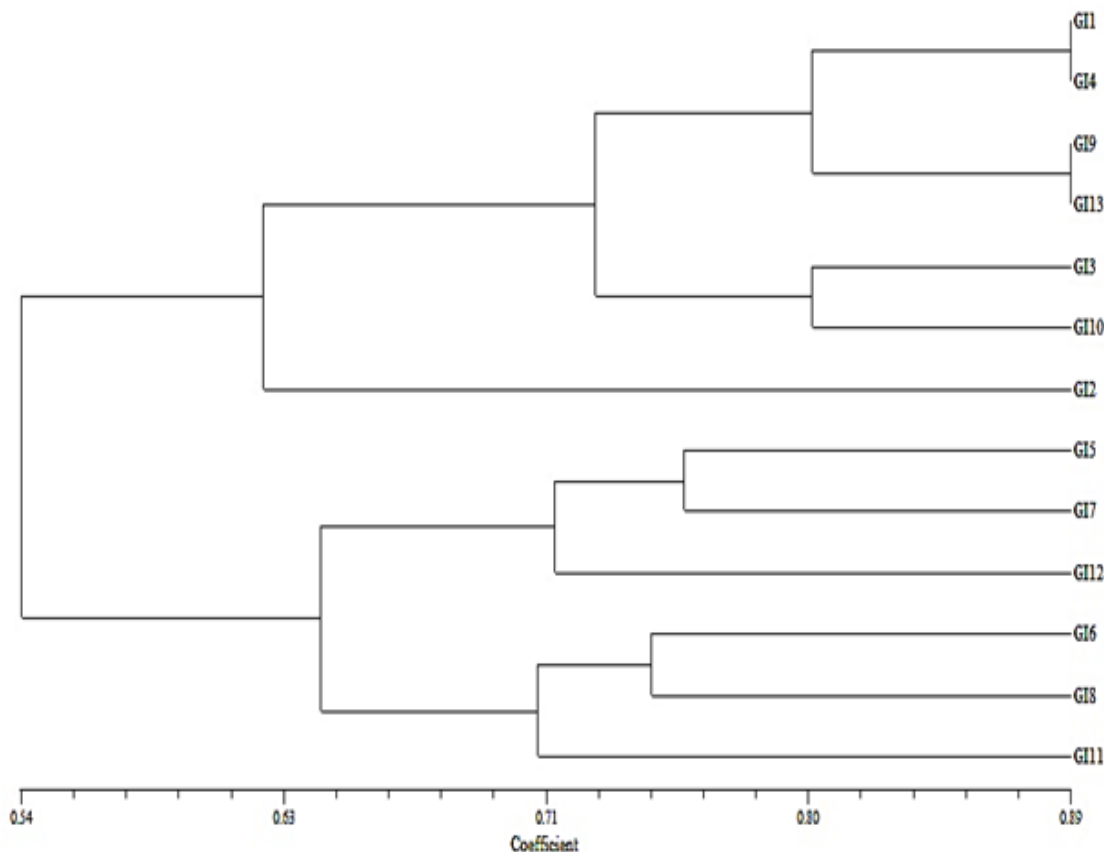


Figure2. UPGMA dendrogram of 13 iron tree genotypes based on 12 random RAPD primers.

Table8. Similarity matrix between genotypes based on SMC similarity.

Row	Genotype	1	2	3	4	5	6	7	8	9	10	11	12	13
1	GI ₁	1												
2	GI ₂	0.65	1											
3	GI ₃	0.76	0.47	1										
4	GI ₄	0.89	0.71	0.65	1									
5	GI ₅	0.57	0.51	0.70	0.68	1								
6	GI ₆	0.33	0.31	0.53	0.40	0.62	1							
7	GI ₇	0.48	0.42	0.64	0.55	0.76	0.73	1						
8	GI ₈	0.34	0.30	0.52	0.39	0.59	0.75	0.72	1					
9	GI ₉	0.89	0.63	0.78	0.80	0.66	0.44	0.53	0.39	1				
10	GI ₁₀	0.78	0.56	0.80	0.69	0.64	0.53	0.64	0.54	0.67	1			
11	GI ₁₁	0.48	0.35	0.68	0.48	0.62	0.71	0.67	0.70	0.53	0.68	1		
12	GI ₁₂	0.64	0.51	0.72	0.68	0.73	0.63	0.69	0.51	0.75	0.59	0.56	1	
13	GI ₁₃	0.80	0.63	0.73	0.71	0.57	0.53	0.51	0.41	0.89	0.71	0.64	0.64	1

Based on the results of similarity matrix, the highest similarity (0.98%) was found between GI₄ with GI₁ genotypes, GI₉ and GI₁₃ and between GI₁ with GI₉. The lowest similarity also was observed between GI₈ and GI₂ and GI₆ and GI₂ genotypes (Table 8). Cluster analysis gained from UPGMA divided the genotypes at 0.6 into two groups. The first group included GI₁, GI₄, GI₉, GI₁₃, GI₃ and GI₁₀ genotypes and the second group was formed with GI₅, GI₉, GI₇, GI₁₂, GI₆, GI₈ and GI₁₁ genotypes. In the first group, GI₁, GI₄, GI₉ and GI₁₃ were not separated by the applied primers (Fig 2).

3.2.3. D-plot Analysis

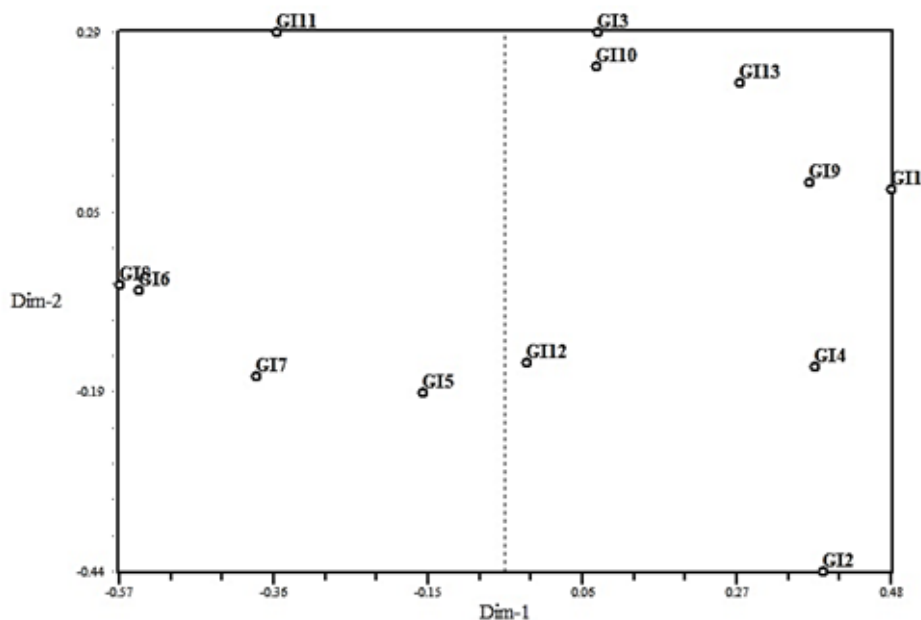


Figure3. Two-dimensional model of the distribution of genotypes using the first two components factors of RAPD data.

This way is used for exhibition of genotypes in bidimensional space based on the effective characters in the first and second factor. The accumulation of one point in plot indicates its genetic similarity. So, based on D- plot analysis, the genotypes which are close to each other in an rear have similar effective traits for factor 1 and factor 2 and are classified in one group. In this study, GI3, GI10, GI13 and GI19 genotypes have higher similarity considering the effective traits in factor 1 and 2 and formed one group. Meanwhile, considering the effective traits in factor 1 and 2, GI11 genotype was at the highest level (positive area) and GI7 genotype was the the lowest level and negative area (Fig 3).

4. DISCUSSION

Seed germination is a mechanism, in which morphological and physiological alterations lead to activation of the embryo. Seed dormancy, on the other hand, is a mechanism by which seeds can inhibit their germination to wait for more favorable conditions (Miransari and Smith 2014). Thus, dormancy and germination are complex phenomena that are controlled by a large number of genes, which are affected by both developmental and environmental factors (Kucera et al. 2005). There are various kinds of dormancy in which physiological dormancy may be decreased by cold or warm stratification or application of hormones (Talos 2001; Zhou et al. 2003; Garcia- Gusano et al. 2004; Rehman et al. 2000; Duan et al. 2004; Fang et al. 2006; El- Refaey, 2014). Based on the present results, combined cold and warm stratification is an efficient way to improve seedling emergence in *Persia parrotia*, as the highest EP was obtained after 42 days warm stratification followed by 50 days cold stratification. Cold and warm stratification had similar effects in species as *Taxus mairei* (Chien et al. 1998), *Stewartia pseudocamellia* (Oleska and Struve 1999), *Rosa taxa* (Alp et al. 2009) and *Styrax japonicas* (Alp et al. 2009). Transverse cut of imbibed seeds of this species indicated that the seeds have a cartilage endosperm most probably having inhibitory compounds, delaying radicle emergence. During stratification, the cartilage endosperm which retards seed germination physically or chemically becomes softened enabling the roots to emerge easily. The existence of thick endosperm has previously reported by Baladan et al. (1995), Bevilacqua et al. (1998), Rascio et al. (1998), Farahmand (1999), and Farahmand and Khosh-Khui (2001) in *Cercis siliquastrum*. Our results showed that after 25 and 50 days of stratification, KNO_3 increased seed germination. KNO_3 has been shown to promote the germination of species such as *Sorbus pohuashanesis*

(Bian et al. 2013) and *Plantago lanceolata* (Osman Sarihan et al. 2005). It is reported that polyamines are involved in the germination process (Matilla 1996). The findings of the present study indicated that putrescine at 5 and 10 mM with 25 and 50 days stratification, increased seedling emergence. This result agrees with Sedaghat and Rahemi (2011), considering the positive effect of putrescine on germination rate and seedling growth of pistachio (*Pistacia vera* L.). Polyamines were increased during seed germination in *Araucaria angustifolia* and *Ocotea odorifera* (Pierruzi et al. 2011). Thus, exogenous application of putrescine may mimic the same role in Persian ironwood.

The present study also indicated that warm stratification could be replaced by exogenously applied putrescine, an indication of possible polyamine increase during cold and warm stratification as previously reported by Matilla (1996). In this study, it was found that after 50 days stratification, emergence was increased by application of BA. The positive effect of BA in this research may directly be related to its mechanism of action or indirectly to its permissive effect on GA₃ (Kucera et al. 2005). Although this claim needs a thorough study tracing and measuring the concentrations of these compounds during stratification and germination. In interaction of chemical treatment with stratification, the highest stem length obtained with putrescine when applied after 75 days of stratification. Application of polyamines (Spd, Spm and Put) on pistachio increased both seedling emergence percentage and emergence rate and seedling growth (Sedaghat and Rahemi 2011). Foliar application of putrescine increased plant height, leaves, number of branches and fresh weight of eggplant (*Solanum melongena* L.) under sandy soil conditions (El-Tohamy et al. 2008). The interaction of effects chemical treatment and stratification showed that the highest shoot and root dry mass were obtained with 3 mM BA and 25 days of stratification. Chapurro et al. (1988) studied effects of BA on peach seeds germination and reported that the highest germination was obtained by 24 hr exposure to 50-100 mg L⁻¹ of BA. Stratification at 5 °C for 20 days led to a marked increase in the cytokinins in *Acer saccharum* (Weed et al. 1973). BA increased plant weight of *Gladiolus* (Ram et al. 2002). BA is a cytokinin known to increase cell division and favor shoot formation. The improvement of shoot and root dry mass, may be the result of enhanced photosynthetic activity by BA. Cytokinins are plant regulators that stimulate cell division, photosynthesis and seed germination (Ram et al. 2002). Cytokinins are also important in the mobilization phenomena of plants and chlorophyll formation in plants (Hare and Van Staden 1997; Fletcher and Mc-Cullagh 1971). It has been proposed that the endosperm is a source of cytokinins needed for promotion of cell division in the embryo. After radicle protrusion, a cytokinin peak is associated with α-amylase accumulation (Kucera et al. 2005). Thus, it appears that BA may activate some genes and the resultant enzymes (such as α-amylase) would ultimately lead to accelerated seedling growth through the activation of related pathways. In general, it seems that morphological, structural and biochemical characters are combined to form a barrier against seed germination in endosperm and embryo of *P. persica*. As the transverse cut in the imbibed seed did not improve seed germination, it appears that the nature of dormancy in this species is affected by both seed reserves (endosperm) and embryo. In previous studies have been showed that the collection time of seeds related to humidity level of the seeds during maturation can be effect on seed dormancy (Hidayati et al. 2001). In regard to the high rainfall and relative humidity in the habitat of parrot tree in Iran, it seems that seed dormancy of parrot no affected humidity of seeds. *P. persica* occurs in the moist deciduous forest region south and south-west of the Caspian Sea. The species grows mainly on lying plains and mountain foothills. Its population decreases as the elevation increases. The optimal condition for its growth is in stations from 125-400m (Andrew 1997). In Northern parts of Iran, the annual precipitation is high (2000 mm in some years) in comparison with other parts of the country. So, it appears that this high rainfall which is combined with warm and cold days have an important role in breaking seed dormancy of this endemic species. So, the positive effect of warm and cold stratification treatments on seed germination and seedling growth is a kind of mimic naturally occurs in Persian parrotia habitat.

The assessment of genetic diversity within and between populations is routinely performed at the molecular level using various laboratory-base techniques such as allozyme or DNA analysis which measure level of variation directly (Mondini et al. 2009). In this research based on cluster analysis, the genotypes were classified into two groups. The researches on this species are scarce. Based on only two morphological studies by Yosefzade et al. (2010) and Sattarian et al. (2011), morphological variations

that were found in leaves of Persian parrotia genotypes, is attributed to elevation, temperature and average rainfall. It is previously concluded that the populations grown in lower latitudes with warmer climate, have lower leaf area. As previous reported (Yosefzadeh et al., 21010, Sattarian et al., 2011), the morphological characteristics of leaves are closely orrelated with climatic conditions. But it appears that the analysis of genetic diversity in *Parrotia persica* based merely on morphological markers is not persuasive. The present study indicated that RAPD markers are better means in this regard and can separate the genotypes of Persian ironwood without the influence of climatic conditions.

In the present research, RAPD markers could separate the genotypes of *Persia parrotia* which are in line with those reported on *Hammamelis* (Marquard et al. 1997). According to Marquard et al. (1997) RAPD markers reasonably discriminated the species with North American origin from the Asiatic species. Based on the present results, the highest resolving power was found with OPD-05 primer and the highest polymorphism was obtained with OPD-05, OPA-10 and OPA-09 primers. Thus, these primers can possibly use for ongoing genetic variation studies in *Persian parrotia*. In this study, 92 bands with the average of 7.66 bands per primer was produced and most produced bands were polymorphic indicating high genetic variation in the studied genotypes. As *Persian parrotia* is a wind- pollinated species, variation is inevitably found in seed-derived genotypes. Meanwhile, this species is naturally grown in several habitats in north of Iran and it is possible that genetic balance has been occurred due to geographical factors. So, this variation needs a comprehensive program evaluating the variation between and among genotypes. Furthermore, *Parrotia persica* is clonally propagated by root suckers in nature. This species has reported as one of the key trees of Caspian-Hyrceanian forests of Iran (Heshmati, 2007). The Hyrcanian forests in Iran and Azerbaijan, together with the Colchic forests of Georgia, are the most important relics of the so-called Arcto- Tertiary forests in western Eurasia and an important biodiversity 'hot spot'. Many tree genera like *Pterocarya*, *Albizzia*, *Parrotia*, or *Gledetsia*, survived the last ice age only in this area (Scharnweber et al., 2007, Andrew, 2007). Because of the fact that most of the temperate deciduous forests in Europe and western Asia are converted into artificial plantations, secondary woodlands or agricultural and urban land, these remnants of primary forests have to be considered as of irreplaceable value (Scharnweber et al. 2007). On the other hand, *Parrotia persica* is a slow growing tree or small shrub and conservation of the present populations and gathering genetic database information for future purposes are of vital importance. Consequently, optimizing protocols for genetic assessment and seed germination to pave the ground for breeding and biotechnological goals, are very decisive.

REFERENCES

- Ahanjan M, Mohana DC, Raveesha KA, Azadbakht M (2007) Antibacterial potential of extracts of leaves of *Parrotia persica*. *Afric J Biotech* 6: 2526- 2528.
- Airi S, Bhat ID, Rawal RS, Dhar U (2009) Variations in seed germination of *Hippophanae salicifolia* with different presoaking treatments. *J Forest Res* 20: 27-30.
- Andrews S (2007) Tree of the Year: *Parrotia*. International Dendrology Society. Yearbook.
- Baldan B, Bonaldo A, Rvitalo N, Meggio F, Profumo P, Mariani P (1995) *Cercis siliquastrum* L.: A comparative study of endosperm and embryo development and resreve accumulation. *Inter J Plant Sci* 156: 181-187
- Bujarska B (2002) Breking of seed dormancy germination and seedling emergence of the common hawthorn (*Crataegus monogyna* Jacq.) *Dendr* 47: 61-70.
- Baninasab B, Rahemi M (2008) The effects of scarification, cold stratification and gibberellic acid treatments on germination of Khokhong seeds. *J Plant Sci* 3: 121- 125.
- Bevilaqua LR, Profumo P, Gastaldo P, Barella P (1998) Cytochemical study of the dormancy-imposing endosperm of *Cercis siliquastrum* L. *Botany* 16: 561-566.
- Bian L, Yang L, Wang J, Shen H (2013) Effects of KNO₃ pretreatment and temperature on seed germination of *Sorbus pohnuashanensis*. *J Forest Res* 24:309-316.
- Brian AO, Struve DK (1999) Germination of *Stewartia pseudocamellia* seeds is promoted by desiccation avoidance, gibberellic acid treatment and warm and cold stratification. *J Environ Hort* 17: 44-46.
- Bryan AA, Seiler JR (1991) Accelerating Fraser fir seedling growth with benzylaminopurine sprays. *HortSci* 26: 389-390.

- Cetnibas M, Koyunncu F (2006) Improving germination of *Prunus avium* L. seeds by gibberellic acid, potassium nitrate and thiourea. Hort Sci 33: 119-123.
- Chaparro JX, Moore GA, Sherman WB (1988) Effect of BA on germination of non stratified peach seed. Acta Hort 254p.
- Chien CT, Huang LLK, Lin TP (1998) Changes in ultrastructure and Abscisic acid levels and response to applied gibberellins in *Taxus mairei* seeds treated with warm and cold stratification. Annals of Bot 81: 41-47.
- Deng, M.B., H.T.Wey and X.Q. Wang (1992). *Shaniodendron*, a new genus of Hammamelidaceae from China. Acta Phy. Taxo. Sinica 30: 57-61.
- Dirr MA (1998). Manual of Woody Landscape Plants. Stipes Publishing, Champaign L.L.C.
- Duan C, Wang B, Liu W (2004) Effects of chemical and physical factors to improve the germination rate of *Echinaceae angustifolia* seeds. Colloids and Surfaces B: Biointerfaces 37: 101-105.
- El-Refaey FA, El-Dengawy A, Hussein AA (2014) The effect of treating persimmon (*Diospyros lotus*) seeds with moist-chilling and growth regulators on seed germination, the subsequent seedling characters and their induced drought tolerance. J Agri Vet Sci 7: 45-53.
- El-Tohamy WA, El-Abagy HM, El-Greadly NHM (2008) Studies on the effect of putrescine, yeast and vitamin C on growth, yield and physiological responses of eggplant (*Solanum melongena* L.) under sandy soil conditions. Aust J Basic Appl Sci 2: 296-300.
- Fang S, Jiayuan W, Zhaoyang W, Zhenxian Z (2006) Methods to break seed dormancy in *Cyclocarya paliurus* (batal) Iljinskaja Sci Hort 110: 305-309.
- Farahmand H, Hashemipour M, Karimi HR, Mohammadi-Nejad G, Bagheri V (2015). Characterization and evaluation of genetic diversity of some old cypress genotypes (*Cupressus sempervirens* L.) in Iran using leaf mineral concentration, biochemical characteristics and SSR markers. Plant Syst. Evol. 301: 761-772.
- Farahmand H (2000) Sexual and Asexual Propagation of Judas tree (*Cercis siliquastrum* L.). M.S.c. Thesis. Department of Horticulture, College of Agriculture, Shiraz University, Shiraz, Iran 65P.
- Farahmand H, Khosh-Khui M (2001) Investigation on improvement of sexual and vegetative propagation of Judas tree (*Cercis siliquastrum* L.) Iranian J Hort Sci Tech 2: 25-38.
- Fletcher RA, Mc-Cullagh D (1971) Cytokinin-induced chlorophyll formation in cucumber cotyledons. Planta 101: 88-90.
- Garcia-Gusano M, Martinez-Gomez P, Dicenta F(2004) Breaking seed dormancy in almond (*Prunus dulcis* D.A. Webb.). Sci Hort 99: 363-370.
- Gilman EF, Watson DG (2014) *Parrotia persica*: Persian Parrotia. Environmental Horticulture Department, Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences (IFAS), University of Florida, Gainesville FL 32611.
- Ghildiyal SK, Sharma CM, Khanduri VP (2009) Effect of pre-soaking and pre-chilling treatments on seed germination of *Pinus roxburghii* provenances from western Himalaya. Indian J Forest Res 20:323-330.
- Grechko VV (2002). Molecular DNA markers in phylogeny and systematics. Russ. J. Genet. 38: 851- 868.
- Hao RM, Wey HT, Liu WG (1996). Floral morphology of *Shaniodendron* (Hammamelidaceae) and its taxonomic significance. J. Plant Res. Envir. 1: 30-35.
- Hare PD, Van Staden J (1997) The molecular basis of cytokinin action. Plant Growth Regul 23:41-78.
- Hector EF, Filner P (1985) Polyamine catabolism in higher plant: Characterization of proline dehydrogenase. Plant Growth Regul 3:277-291.
- Heshmati GA (2007) Vegetation characteristics of four ecological zones of Iran. Inter J Plant Product 2: 215-224.
- Hidayati SN, Baskin JM, Baskin CC (2001) Dormancy-breaking and germination requirements for seeds of *Symphoricarpos orbiculatus* (Caprifoliaceae). Amer J Bot 8:1414-1451.
- Karimi HR, Kafkas S, Zamani Z, Ebadi A, Fatahi MR (2009) Genetic relationships among species and cultivars of *Pistacia* using AFLP marker. Plant Syst. Evol. 279:21–28
- Karimi HR, Sadeghi-Seresht E (2015). Investigation of genetic nature and diversity of Banebaghi as genotype of *Pistacia* genus by using morphological characteristics and RAPD marker. Plant Cell Biot. Mol. Biol. In press
- Khalil RY, Al-Eisawi DM (1998) Seed germination of amygdalus of Arabica olive; As influenced by stratification by stratification and certain plant bioregulators. Acta Hort 517.

- Li J, Tredici PT (2006) The chinese parrotia: A sibling species of the *Persian parrotia*. *Arnoldia J Forest Res* 18: 57-62.
- Marquard RD, Davis ED, Stowe EL (1997). Genetic diversity among witchhazel cultivars based on Randomly Amplified Polymorphic DNA markers. *J. Amer. Soc. Hort. Sci.* 122:529-534
- Matilla AJ (1996) Polyamines and seed germination. *Seed Sci Res*, 6: 81-93
- Metreveli M, Bregvadze M (2007) Peculiarities of propagation of species of Hamamelidaceae Lindl. Family. *Bullet Ger Nat Acad Sci*, 175.
- Miransari M, Smith D L (2014) Plant hormones and seed germination. *Envir Exper Bot* 99:110-121.
- Mozaffarian VA (2003) A Dictionary of Iranian Plant Names (Latin, English, Persian). Farhang Mosaer Publishers. Tehran, Iran. 671p. (In Persian).
- Mozaffarian VA (2005) Trees and Shrubs of Iran. Farhang Mosaer Publishers. Tehran, Iran. 990p. (In Persian).
- Mondini L, Noorani A, Pagnotta MA (2009). Assessing plant genetic diversity by molecular tools. *Diversity* 1: 19-35
- Oleska BA, Struve DK (1999) Germination of *Stewartia pseudocamelia* seeds is promoted by desiccation avoidance, gibberellic acid treatment and warm and cold stratification. *J Envir Hort* 17: 44-46
- Osman Sarihan E, Ipek A, Mahmood K (2005) Role of GA₃ and KNO₃ in improving the frequency of seed germination in *Plantago lanceolata* L. *Pakistan J Bot* 37: 883-887.
- Pieruzzi FP, Dias LLC, Balbuena TS, Santa-Catarina C, dos Santos ALW, Floh EIS (2011) Polyamines, IAA and ABA during germination in two recalcitrant seeds: *Araucaria angustifolia* (Gymnosperm) and *Ocotea odorifera* (angiosperm). *Anal of Bot* 133: 1-9.
- Rahemi M, Baninasab B (2000) Effect of gibberellic acid on seedling growth in two wild species of pistachio. *J Hort Sci Biotech* 75: 336-339
- Ram, R., Muknerjee, D., Manujas, S (2002) Plant growth regulators affect the development of both corms and cormlets in *Gladiolous*. *Hort Sci*, 37: 343-344
- Rehman S, Park IH (2000) Effects of stratification, GA₃ and chilling on germination of goldenrain-tree (*Koelreuteria paniculata* Laxm.) seeds. *Sci Hort* 85: 319- 324.
- Roh MS, Bentz JA, Wang P, Li H, Koshioka M (2004) Maturity and temperature stratification affect the germination of *Styrax japonicus* seeds. *J Hort Sci Biotech* 79: 645-651.
- Sabeti HA (1994) Forest, Trees and Shrubs of Iran. Yazd University Press. Yazd, Iran. 806p. (In Persian).
- Sattarian A, Akbarian MR, Zarafshar M, Bruschi P, Fayyaz P (2011) Phenotypic variation and leaf fluctuating asymmetry in natural population of *Parrotia persica* (Hamamelidaceae), an endemic species from the Hyrcanian forest (Iran). *Acta Bot Mex* 97: 65-81.
- Salim Azad Md, Mizanur Rahman M, Shamin Hasan MD, Abdul Matin MD (2011) Effect of different pre-sowing treatments on seed germination percentage and growth performance of *Acacia auriculiformis*. *J Forest Res* 22:183-188
- Salim Azad Md, Al-Musa MZ, Abdul Matin Md (2010) Effects of pre-sowing treatments on seed germination of *Melia azedarach*. *J Forest Res* 21: 193-196
- Scharnweber T, Rietschel M, Manthey M (2007) Degradation stages of the Hyrcanian forests in southern Azerbaijan. *Archiv fur Naturschutz und Landschaftsforschung Juni 2007* Institute of Botany und landscape Ecology, Greifswald University, Germany
- Sedaghat S, Rahemi M (2011) Effect of pre-soaking seeds in polyamines on seed germination and seedling growth of *Pistacia vera* L. cv. Ghazvini. *Inter J Nut. Relat Sci* 2: 7-14.
- Shanjani P, Pule L, Khavri-Nejad RA, Gomory D, Sagheb-Talebi K (2002). Genetic diversity of oriental Beech (*Fagus orientalis* Lipsky.) forest over the hHyrcanian zone. *Forest Gen.* 9 297-308
- Strik WA., Gold JD, Novak, O, Strand M, van Staden J (2005) Changes in endogenous cytokinins during germination and seedling establishment of *Tagetes minuta* L. *Plant Growth Regul* 47: 1-7.
- Takos IA (2001) Seed dormancy in bay laurel (*Laurus nobilis* L.). *New Forests* 21: 105-114.
- Tzortzakis NG (2009) Effect of pre-sowing treatment on seed germination and seedling vigour in endive and chicory. *HortSci* 36: 117-125.

Effects Chemical Treatments and Stratification on Seedlings Emergence of Persian Parrotia (*Parrotia Persica* (DC.) and Assessment of Genetic Diversity in its Seedlings

- Rohle JF (2004) NTSYS-pc: 2.11 numerical taxonomy and multivariate analysis system, version 2.11. Exeter Publishing, Ltd, New York.
- Yousefzadeh H, Tabari M, Akbarinia M, Akbarian MR, Bussoti F. 2010. Morphological plasticity of *Parrotia persica* leaves in eastern of Hyrcanain forests (Iran) is related to altitude. Nord. J. Bot. 28(3): 344-349
- Wada S, Reed BM (2011) Standardizing germination protocols for diverse raspberry and blackberry species. Sci Hort 132: 42-49.
- Weeb DP, Van Staden J, Wareing PE (1972) Seed dormancy in *Acer* changes in endogenous cytokinins, gibberellins germination inhibitors during the breaking of dormancy *Acer saccharum* marsh. J Exper Bot 24: 105-116.
- Zhi-yun Z, An-ming L (1995) Hamamelidaceae: Geographical distribution, fossil history and origin. Acta Physiol Taxo Sci 33: 313-339.
- Zhou L, Wu J, Wang S (2003) Low-temperature stratification strategies and growth regulators for rapid induction of *Paris polyphylla* var. *yunnanensis* seed germination. Plant Growth Regul 41: 179-183
- Zhou Z, Crepet WL, Nixon KC (2001) The earliest fossil evidence of the Hamamelidaceae: Late Cretaceous (Turonian) inflorescences and fruits of Altingioideae. Amer J Bot 88: 753- 766.

Citation: Hamid Reza Karimi, et al., "Effects Chemical Treatments and Stratification on Seedlings Emergence of Persian Parrotia (*Parrotia Persica* (DC.) and Assessment of Genetic Diversity in its Seedlings" *International Journal of Advanced Research in Botany*, vol. 4, no. 3, p. 1-15, 2018. <http://dx.doi.org/10.20431/2454-9444.0403001>

Copyright: © 2018 Authors. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.